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COMPARATIVE STUDY TO INVESTIGATE DIFFERENT ENZYME SOURCES ON THE ENERGY UTILIZATION IN BROILER DIETS

BY

Wesam Sadeq Abdallah Abu Suliman

**A Thesis Submitted As Partial Fulfillment of the
Requirements for the Master Degree**

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**Submitted in Partial Fulfillment of the Requirements for the Degree of
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In

**Agriculture Science (Poultry Nutrition)
Department of Animal and Poultry Production
Faculty of Agriculture,
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Supervision Sheet

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1- INTRODUCTION

The application of feed enzymes to poultry diets for the enhancement of nutrient availability has been practiced for long time ago. Early research conducted on feed enzymes in poultry nutrition focused on non-starch polysaccharide (NSPs) degrading enzymes, specifically xylanase and β -glucanase, in diets containing wheat, rye and barley (**Choct, 2006**). Poultry do not synthesize the enzymes necessary for the digestion of cellulose, β -glucans, arabinoxylans or pectin. Non-starch polysaccharides are complex sugars that occur in the cell walls of cereal grains (**Bedford and Schulze, 1998**). It includes arabinoxylans in rye and wheat (**Beg et al., 2001**). Enzymes such as xylanase and β -glucanase, have the ability the break down these structural polysaccharides, making the nutrients available to the animal (**Bedford, 2000, and Choct, 2006**).

The application of exogenous fibrolytic enzymes in the poultry industry have been used to neutralise the effects of NSPs present in wheat, barley, rye and oats and this has proven to be a successful means of improving broiler production performance (**Choct et al., 1995, Brufau et al., 2001, Engberg et al., 2004, Cowieson et al., 2005, Scott, 2005, Wang et al., 2005, and Pettersson and Åman, 2007**). A summary of some published results presented by **Cowieson (2005)** reveals that the use of a combination of xylanase, protease, and amylase resulted in significant improvements in feed:gain (from 0.8 to 10.5%) and BW gain (from 1.9 to 6.9%). The same enzyme combination increased AME by 3% and nitrogen retention by 11.7% of both adequate and reduced energy and amino acid diets (**Cowieson and Ravindran, 2008**).

The effects of phytase in poultry have recently been extensively reviewed by **Selle and Ravindran (2007)**. They concluded that phytase increases phosphorus retention and tibia ash, but it has also positive effects on weight gain, feed intake, nitrogen retention, feed gain ratio, apparent metabolisable energy, and Ca retention.

Studies have shown that supplementing enzymes to broiler could increase the AMEn of the diets (**Fuente *et al.*, 1995, Huyghebaert, 1997, Choct *et al.*, 1999, and Cowieson and Ravindran, 2008**) which could participate to decrease the cost of the diets

The objective of this study was to compare production performance of broilers receiving diets containing optimum or low metabolisable energy levels supplemented with a phytase, enzyme mixture, or a combination of the phytase and the enzyme mixture supplemented to wheat based diets.

2. REVIEW OF LITERATURE

2.1. Dietary energy level

Cheng *et al.*, (1997) reported that male broilers fed high energy diet (3250 kcal/kg) across temperatures ranged from 21.1°C-35°C had significantly improved feed conversion compared to broilers fed with 3000 kcal ME/kg diet.

El-Husseini *et al.*, (2002) reported that decreasing the dietary metabolizable energy from 3200 to 3000 kcal/kg reduced significantly the body weight gain of the broilers.

Maiorka *et al.*, (2004) showed that broilers fed the diet containing 3,200 kcal ME/kg had higher weight gain (P=0.015), better feed conversion (P=0.001), and higher abdominal fat deposition (P=0.001) as compared to those fed the diet containing 2,900 kcal ME/kg.

Downs *et al.*, (2006) reported that decreasing dietary ME level about 100 kcal/kg during each growth phase resulted in comparable live body weight for broilers. While feed conversion ratio was significantly improved with increasing dietary ME level. Carcass yield and abdominal fat percentages were not significantly affected by dietary ME level.

El Tazi *et al.*, (2009) studied the effect of dietary metabolizable energy levels (3000 *vs.* and 3200 Kcal/kg) on body weight, feed efficiency and dressing percentage was not significant, while the feed intake was decreased significantly by increasing the level of metabolizable energy.

Golian *et al.*, (2010) reported that live body weight and feed conversion ratio were improved as the dietary energy level was increased (2900, 3000, 3100 and 3200 kcal ME/ kg diet).

Jafarnejad and Sadegh (2011) reported no significant difference on live weight of broilers fed 3200 vs. 3000 kcal /kg diet.

2.2. Enzymes

Monogastrics do not synthesize enzymes capable of hydrolyzing cereal structural carbohydrates, but do possess microorganisms in the large intestine and caecum that have this capability. Enrichment of their digestive system with exogenous enzymes is of utmost importance in practical feeding especially in young broiler chicks that have a relatively low level of posterior small intestine fermentation (**Engberg et al., 2004,** and **Meng et al., 2004,** and **2005**). It is hypothesized that the response to enzyme supplementation of cereal-based diets is significantly higher in younger compared to older animals, as young chicks are sensitive to the water-soluble and highly viscose carbohydrates (**Boros et al., 1998**).

2.2.1. Non starch polysaccharides

Non-starch polysaccharides mostly present in raw materials used for poultry diets are pectins, cellulose, mixed-linked β -glucans and arabinoxylans (**Parsippany, 2008**). Non-starch polysaccharides (NSPs) can be divided into two groups, soluble and insoluble NSPs (**Castanon et al, 1997**). Starch can function as a major nutritional component of the diet, but a number of NSPs can have negative effects on poultry. Poultry cannot digest the insoluble NSPs, only soluble NSPs have the potential to be digested by poultry (**Parsippany, 2008**). According to **Carre et al., (1995)** the degradation of soluble NSP can be as high as 80-90 %, while the insoluble NSPs remain un-digested.

After the ingestion of NSPs, the NSPs bind to the water in the intestinal tract of the bird due to the water-holding capacity of the soluble fibres and this leads to an increase in the viscosity of the digestant (**Classen, 1996**). This increase in the intestinal viscosity leads to a decreased rate of digestion, which in turn leads to a reduction in performance (**Bedford, 1993, Classen, 1996, and Chesson, 2001**). Also, NSPs leads to an increase in the thickness of the intestinal contents and result in gelatinous droppings (sticky droppings). The moisture content of litter is increased by the sticky droppings and this leads to wet litter. (**Selinger *et al.*, 1996, Bedford and Schulze, 1998, Bhat, 2000, Hetland *et al.*, 2004, and Nnenna *et al.*, 2006**).

Depolymerisation of these NSPs requires specific enzymes; these enzymes are specific to the main and side chain structure of the NSPs (**Bedford and Schulze, 1998, Andersson *et al.*, 2003, Bhat, 2000, and Botha, 2011**).

The major NSPs found in wheat are arabinoxylans and it is the major contributor to the soluble NSPs fraction of wheat and barley (**Beg *et al.*, 2001, and Chesson, 2001**). The endosperm and the bran of wheat is the major source of soluble arabinoxylans. Arabinoxylan from the bran contributes 33.3 % to the total soluble arabinoxylan content of wheat and bran contributes 64.2 % (**Chesson, 2001**). The NSPs content of wheat according to **Knudsen (1997)** is 119 g/kg, 25 g/kg of the total NSPs are soluble. The largest contributors to the soluble non-cellulosic polysaccharide fraction of wheat are xylose, arabinose and glucose at 9 g/kg, 7 g/kg and 4 g/kg respectively (**Knudsen, 1997, and Fourie, 2007**). NSPs of concern in wheat are the insoluble non-cellulosic polysaccharide and cellulose. The insoluble non-cellulosic polysaccharide content of wheat is 74 g/kg of the DM content of wheat, 62 % of the total

NSP content of wheat. Cellulose contributes 17 % to the total NSPs content of wheat and 2 % of the DM content of wheat (**Knudsen, 1997**).

Addition of the appropriate enzyme diminishes these constraints and allows digestion to occur more rapidly and completely. Exogenous enzyme is, therefore, likely to supplement the digestive capacity of the younger bird, and again the response to such will be more apparent with poorly digested compared with highly digestible diets. It is likely that as a bird age and their digestive ability increases, as does their microbial population, the effect of exogenous enzymes is more and more mediated through the microbial route (**Bedford, 2000**).

The beneficial effects of exogenous enzyme supplementation depend on a number of factors. These factors include the dietary components, processing of the diet and the type of enzymes used (**Acamovic, 2001**).

2.2.2. Carbohydrases

Carbohydrases' enzymes may hydrolyze the aleurone layer thus reducing barrier to nutrient digestibility and thus enhance the utilization of the feedstuffs. All cereals have aleurone layer which shields the starchy endosperm thus presenting a first barrier to digestibility and nutrient availability (**Oluyinka and Adeola, 2008**).

Wheat arabinoxylans act as anti-nutrient by increasing digesta viscosity thus impair nutrients utilization (**Evers *et al.*, 1999**). **Bedford and Classen (1992)** demonstrated that improvement in feed conversion ratio and weight gain in broiler chickens given wheat-based diets supplemented with xylanase was significantly correlated with a reduction in viscosity of the jejunum and ileum content. Xylanase enzymes

hydrolyse the soluble arabinoxylans and thus prevent high intestinal viscosities and improve nutrient absorption (**Van der Klis *et al.*, 1995**).

It is worth noting that the beneficial effects of xylanase addition to a wheat-based broiler diet were slightly dependent on wheat content since increased inclusion level of wheat induced higher digesta viscosity (**Veldman and Vahl, 1994**). Also, the beneficial effect of the use of exogenous enzymes may depend on the wheat cultivar used (**Gutierrez *et al.*, 2008**).

Marsman *et al.*, (1997) conducted a study where a cell wall degrading enzyme preparation (EnergexTM) was added to a maize-SBM based diet for broilers. The carbohydrase preparation consisted of cellulase, hemicellulase and pectinase. Their results indicated that body weight gain, feed intake and feed conversion ratio were not affected by the enzyme supplementation.

Steenfeldt *et al.*, (1998) reported that supplementation with xylanase enzymes containing endoxylanase 350 U/g or endoxylanase 400 U/g had a significant effect on broiler performance during the first 3 weeks, and from 21-42 days of age compared to control group. Feed conversion efficiency was in general similarly affected.

Zanella *et al.*, (1999) supplemented a maize-SBM based diet with a commercial enzyme preparation, Avizyme[®] 1500, it was added to the diet at 0.1 %. Avizyme[®] 1500 consists of 800 µ/g xylanase from the fungus *Trichoderma longibrachiatum*, 6,000 µ/g protease from the gram-positive bacterium *Bacillus subtilis*, and 2,000 µ/g amylase from the gram-positive bacterium *Bacillus amyloliquifaciens*. Enzyme supplementation also led to the significant increase of 1.9 % in live

weight and improvement of 2.2 % in feed conversion ratio. Abdominal fat content of broilers was not significantly affected by treatment.

Saleh et al., (2005) revealed that the enzyme preparation had no effect on the abdominal fat content of broilers. The enzyme preparation used for this study was a pure cellulase (1, 4-(1,3:1,4)-b-D-Glucan 4- glucano-hydrolase) obtained from the fungus *Trichoderma viride*. The addition of the pure cellulose to the maize-SBM based diet had no effect on body weight gain and feed conversion ratio.

Pettersson and Aman (2007) tested the addition of an enzyme cocktail, consisting of xylanase and β -glucanase to broiler diet containing rye and wheat. The addition of the enzyme cocktail resulted in a significant increase in body-weight and feed intake.

Oluyinka and Adeola (2008) reported wheat-based diets benefit from supplementation of microbial xylanase because of the high level of arabinoxylans in wheat.

Owens et al., (2008) compared between different xylanses sources and reported improved total live weight gain (+12%) and feed conversion ratio (+9%) for all enzyme-supplemented diets compared to the control fed broilers. They added that Allzyme, the preparation of endo-1, 4-beta-xylanase with an activity level of 600 U/g, alone was as effective as any of the combinations with organic acids or mannan oligosaccharides.

Lö et al., (2009) found that broiler diet supplemented with Avizyme 1302 (300 g/ton feed to supply 2000 xylanase units/kg diet) had no significant on broiler performance.

Kwakernaaka et al., (2011) added endo-xylanase (0, 100 and 200 U/kg) to turkey diets. Dietary supplementation at 200 U/kg

significantly improved body weight gain and feed conversion ratio over a 16-week period.

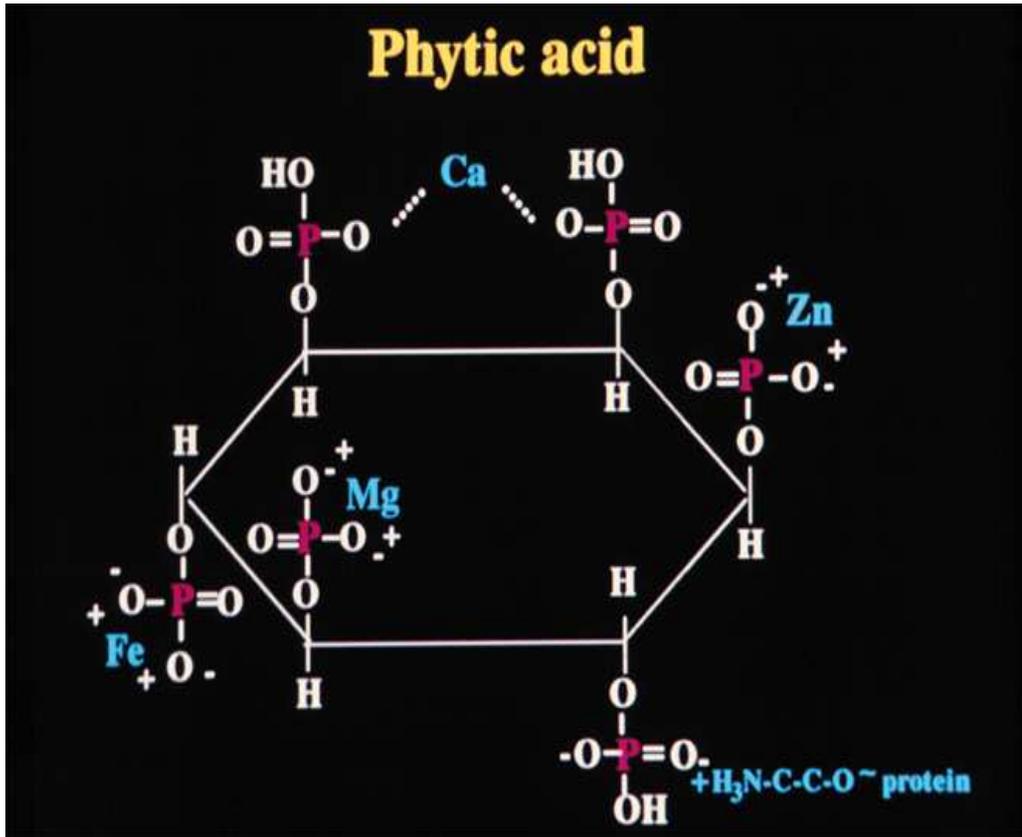
2.2.3. Phytase

2.2.3.1. Phytate

Phytate, the mixed salt of phytic acid (myoinositol hexaphosphate, IP6), is a ubiquitous component of plant-source feed ingredients; consequently, it is invariably present in practical broiler diets at levels ranged from 2.5 to 4 g/phytate-P/kg (**Ravindran, 1995**). It has been estimated by **Lott *et al.*, (2000)** that the total global harvest of barley, maize, sorghum, wheat, cottonseed, rapeseed and soyabean, which are considered key feed ingredients for poultry, contains approximately 16 million tones of phytate. Phytate is predominantly present as the myoinositol hexaphosphate ester (IP6) (**Kasim and Edwards, 1998**) coupled with magnesium (Mg) and potassium (K) as a mineral–phytate complex. Indeed, **Lott *et al.*, (2000)** proposed that the model substrate is one in which IP6 is complexed with three Mg²⁺ and six K⁺ ions.

The majority of P in plant-source feed ingredients is present as phytate-P, as the phytate-P proportion of total P ranges from 60% (soyabean meal) to 80% (rice bran). That phytate-P which is only partially utilised by poultry is reflected in the poor bio-availability of P. The dietary phytate concentration should govern the magnitude of responses to microbial phytase and such indications have been reported in poultry (**Cabahug *et al.*, 1999, Ravindran *et al.*, 2000, and 2006**).

Phytic acid



Edwards (1991)

2.2.3.2. Phytic acid content in feedstuffs

Data presented in Table (1) declared the phytic acid contents of feedstuffs and its intrinsic phytase activity as well as optima pH and temperature.

Table (1): Phytin content in some common feedstuffs used in dietary formulation for poultry and their intrinsic phytase activity (Godoy *et al.*, 2005¹, Kornegay *et al.*, 1996², and Greiner and Konietzny, 2006).

Feedstuffs	Phytate		Phytase activity, U/ kg	Optimum ³	
	%	% of total P		pH	Temperature °C
Maize ¹	0.17	73	24	4.8	55
Sorghum ¹	0.17	66	24	NR	NR
Wheat ¹	0.18	55	1193	5-6	45-50
Wheat bran ¹	0.63	69	2957	NR	NR
Triticale ²			1500	NR	NR
Barley ²	0.27	64	582	5-6	45-55
Oats ²	0.29	67	40	5.0	38
Rice polishing ¹	1.13	72	134	NR	NR
Soybean	0.37	65	62	4.5-5.0	55-58
Canola meal ²	0.70	59	16	NR	NR
Sunflower meal ²	0.89	77	60	NR	NR
Peanut meal ²	0.48	80	3	NR	NR
Cottonseed meal ¹	0.84	63	36	NR	NR
Palm oil meal ¹	0.29	57	34	NR	NR
Coconut meal ¹	0.24	56	37	NR	NR
Brewery grains ¹	0.30	24	39	NR	NR
Sesame seed meal ⁴	3.57	NR			

NR, not reported, ⁴ Eardman (1979).

2.2.3.3. Phytase and broiler performance

Qian *et al.*, (1996) reported that live weight gain of broilers was improved by the addition of 400, 600, and 800 FTU/kg to 0.34%, 0.27%, and 0.20% non-phytate phosphorus diets, respectively.

El-Saady (1999) reported that 750 FTU/kg broiler diet improved significantly live weight gain, feed intake and feed conversion ratio, but had no effect on dressing % compared to the control.

Cabahug *et al.*, (1999) revealed that increasing dietary phytic acid from 10.4 to 15.7 g/kg (from rice pollard) negatively impacted live body weight gain, feed intake, and feed conversion ratio, and these adverse effects were partially overcome by phytase addition at 400 and 800 U/kg diet. The improvements occurred in broilers live body weight gain and feed conversion ratio by phytase were greater in high-phytic acid containing diets.

Raw *et al.*, (1999) found that phytase addition at 250 and 500 U/kg diet improved live body weight gain and feed conversion ratio of broilers.

El-Deeb *et al.*, (2000) observed that microbial phytase (Natuphos[®]) addition at 500 or 1000 U/kg of broiler diets significantly increased live weight gain, but had no effect on feed intake or feed conversion ratio.

Abd El-Samee (2002) reported that when diets containing medium level of crude protein and fungal phytase (750 U/kg) were fed, the broiler growth performance was improved significantly compared to the control group. Moreover, results obtained showed no significant effect on carcass characteristics of 7 weeks age that could be attributed to phytase supplementation, irrespective of protein level.

Balasubramanian et al., (2002) noticed that supplementation of different levels of fungal phytase at 500, 750 and 1000 U/kg diet of low available P level (0.3%) significantly increased live body weight gain and improved feed conversion ratio.

Qota et al., (2002) reported that phytase addition at 500 U/kg diet contained 10% linseed cake did not affect live weight gain, and feed conversion ratio. Also, they found that phytase addition at 500 U/kg of 10% soaked linseed cake-diet had no effect on plasma total protein, triglycerides, and cholesterol. Moreover, it did not significantly affect dressing or giblets %.

Abdo (2004) studied the effect of phytase addition (0.0, 500, and 1000 U/kg diet contained 0, 25 and 50% level of *Nigella sativa* seed meal to substitute soybean meal. Phytase addition at 500 or 1000 U/kg diet with 25% *Nigella sativa* seed meal resulted in the highest live body weight gain compared to the control group. Phytase had no significant effect on carcass percentage, or liver function (AST activity).

Aggoor et al. (2004) found that phytase addition at 600 U/kg diet containing rapeseed meal increased live weight gain and improved feed conversion ratio of broilers.

Hassanabadi et al. (2004) found that phytase at the rate of 250, 500, 750 and 1000 U/kg diet improved live weight gain, feed intake, and feed conversion ratio of broilers.

Onyango et al., (2005) found that *Escherichia coli* phytase at 500 or 1000 U/kg of diet increased significantly live weight gain, and feed intake of broilers.

Pillai et al., (2006) observed that *Escherichia coli* phytase addition at 250, 500, and 1000 U/kg increased live weight gain and carcass yield of broilers.

Selim (2007) reported that either Natuphos[®] (Phytase-3) or Phyzyme (Phytase-6) at 500 U/kg diet decreased significantly growth of Sasso chicks from 35 to 56 day of age compared to the un-supplemented control. However, Natuphos[®] increased significantly growth of chicks from 56 to 64 d of age compared to the un-supplemented control and Phyzyme supplemented groups. Meanwhile, difference was insignificant from one-day old to 35 and one-day old to 64 day of age.

2.2.4. Integration between carbohydrases and phytase enzymes

The use of an enzyme complex containing carbohydrases and phytase should allow the reduction of dietary available P and energy, as well as crude protein, amino acids, and Ca concentrations in feed. Some attempts have been made to assess the magnitude of dietary nutrient reduction, which can be compensated by xylanase, amylase, protease, and phytase to maintain performance to that of the positive control, containing adequate nutrient concentrations (**Cowieson et al., 2006**).

Carbohydrases, which are able to break down the cell wall NSPs matrix, can facilitate the access of phytase to phytate molecule (**Olukosi et al., 2007**), supporting the hypothesis that the use of a combination of carbohydrases and phytase can fully strengthen their effects.

Ravindran et al., (1999) studied the response of broiler chicks to microbial phytase (Natuphos[®] 5000) added at the rate of 120 g/ton, individually and in combination with glucanase preparations with

predominantly xylanase (Natugrain Blend) and glucanase (Natugrain) enzymes. Performance of broilers fed a wheat-based diet was not influenced by the addition of individual enzymes, but increasing inclusion levels of the xylanase plus phytase combination linearly improved live weight gain and feed efficiency.

Francesch and Geraert (2009) reported improved weight gain and feed intake and energy utilization in broilers fed corn-soybean diets deficient in available phosphorus. This might indicate that the first limiting nutrient was phosphorus and once the phosphorus deficiency was overcome by phytase and feed intake was restored, the carbohydrases could increase the nutritive value of diet compensating for the reduction in AME (+ 2.8%).

2.3. Interaction between dietary energy level and enzyme supplementation

The main objective of the use of NSPs-degrading enzymes in wheat and barley based diets is to increase the apparent metabolisable energy of the wheat (**Botha, 2011**). The variability of the apparent metabolisable energy content of wheat is related to the anti-nutritional effects of the NSPs present in the wheat and barley (**Choct *et al.*, 1995, 1996, and 1999**).

Choct *et al.*, (1995) tested the supplementation of fibrolytic enzymes to broiler diets containing low apparent metabolisable wheat. Their enzyme cocktail was added at 1 g per kg of feed and consisted of xylanase, β -glucanase and pectinase. A 24.3 % increase in apparent metabolisable energy was observed when the enzyme cocktail was added to the low apparent metabolisable energy wheat diet. The most interesting

observation in this trail was the improvement of 34.1 % in the feed conversion efficiency. The most important characteristic of low apparent metabolisable wheat is the incomplete starch digestion with the excretion of starch in the faeces as a consequence. However, starch digestion in the small intestine of broilers is increased with enzyme supplementation.

Fuente *et al.*, (1995) found that enzyme mixture addition significantly increased the AMEn of diets in 30-d-old chicks, but did not see any increase in 10-d-old chicks.

Huyghebaert (1997) indicated that adding multi-enzyme preparations to a diet containing 25 or 50% wheat increased diet metabolizable energy by 4.9 and 9.4%, respectively.

Choct *et al.*, (1999) revealed that the use of xylanase significantly increased the AMEn of the wheat and starch digestibility in the jejunum and ileum of the bird. AMEn increased from 13.7 to 14.5 MJ/kg DM intake.

Zanella *et al.* (1999) found that a combination of xylanase, protease, and α -amylase allowed restoring a reduction of 3.4% in dietary AME in broilers fed corn-soybean meal diet.

Marron *et al.*, (2001) also used xylanase for supplementing a wheat based broiler diet and they also found that enzyme supplementation increased AMEn and starch digestibility.

Kocher *et al.*, (2003) found that the combined addition of pectinase, protease, and amylase significantly improved AMEn when added to a corn soybean meal basal diet with lower energy and protein levels.

Cowieson and Ravindran (2008) reported that a combination of xylanase, amylase, and protease was able to increase the availability of energy by 3% for broiler chicks.

El Tazi *et al.*, (2009) reported that studied the interaction between the dietary microbial phytase (zero *vs.* 500 U/kg diet) and metabolisable energy (3000 *vs.* and 3200 kcal/kg) did not significantly affect the weight gain, feed intake, feed efficiency and dressing percentage of broilers.

3. MATERIALS AND METHODS

The growth trial of the present study was conducted at a private sector poultry farm at Beit-Lahia, North Governorate, Palestine during the period of March to May 2010, while, slaughter test and blood serum measurements were done at Faculty of Agriculture-Al-Azhar University, Gaza strip.

The work aimed at comparative study to investigate different enzyme sources on the energy utilization in broilers fed wheat-base diets.

3.1. Experimental design

Three hundred and twenty one-day-old unsexed Cobb chicks were randomly distributed between 8 experimental groups of 40 chicks, each to study to investigate the effect of supplementing different enzyme sources (phytase *vs.* enzyme mixture) to different dietary metabolisable energy levels (optimum *vs.* low) by broilers fed wheat-based diets on growth performance, carcass traits and some blood serum constituents. Diets were as follows:

Treat. 1: Diet containing the ideal ME energy level without enzyme supplement (control).

Treat. 2: Control + Avizyme 1500 (100 g/100 kg)

Treat. 3: Control + Phyzyme XP TPT 10000 (50g /100 kg)

Treat.4: Control + Avizyme 1500 (50 g) + Phyzyme XP TPT 10000 (25 g /100 kg)

Treat. 5: Diet containing a low-ME (low 100 kcal/kg diet) without enzyme supplement.

Treat. 6: Low-ME diet + Avizyme 1500 (100 g / 100 kg)

Treat.7: Low-ME diet + Phyzyme XP TPT 10000 (50g /100 kg)

Treat. 8: Low-ME diet + Avizyme 1500 (50 g) +

Phyzyme XP TPT 10000 (25g / 100 kg)

3.2. Experimental diets

Throughout the 6 week growth trial, broilers were fed one phase-diet. Composition and calculated analyses of the basal experimental diet were calculated according to **NRC (1994)** are presented in Table 2.

3.3. Management

Feed, light, and water were continuously made available from 1st day old up till 42 days old. Common management practice was used for brooding and rearing. Vaccination and medical care were applied according to the common veterinary practice for broiler chicks. The medical care applied include vaccination against New-Castle disease three times, first by Hitchner B1 strain at the 6th day of age and twice with Lasota strain in drinking water at 18 and 35 days of age, respectively. Chicks were also vaccinated against infectious bursal disease (Gumboro) in drinking water, once at 12 and the other 22 days of age. Feed consumption and live body weight for all experimental groups were recorded on a replicate base weekly through the feeding period.

Table (2): Formulation and nutrient composition of the basal diet

Ingredients	Basal diet %
Yellow corn	36.0
Wheat	20.5
Soybean meal(44%)	36.0
Dry-fat	3.8
Limestone	1.5
Di calcium phosphate	0.10
Nacl	0.35
Vitamins & minerals premix*	0.30
DL- methionine	0.1
L-lysine	0.05
Total	100.0
Chemical composition	
Cp	21.63
ME (kcal/kg)	2945
EE	4.59
Ca	1.00
P	0.46
Lysine	1.10
methionine	0.55

*Vit. & Min. Premix supplied per 1 kg of diet: vit. A.,12000 IU; Vit.D3, 220 IU; Vit. E, 10 mg; Vit.K3, 2 mg; Vit.B1, 1 mg; Vit.B2, 4 mg; Vit.B6, 1.5 mg; Vit.B12, 10 µg; Vit.B3, 20 mg; Vit.B5, 10 mg; Vit.B9, 1 mg; Vit.B8, 50 µg; Choline chloride, 500 mg; Cu, 10 mg; I, 10 mg; Fe, 30 mg; Mn, 55 mg; Zn, 850 mg; Se, 0.1 mg and Co, 0.25 mg.

Wheat bran was included at the rate of 6% to diets 5 up to 8, in order to decrease the energy density of the basal diet by about 100 kcal/kg.

Avizyme 1500 contains xylanase derived from genetically modified *Trichoderma longibrachiatum*, protease derived from genetically modified *Bacillus subtilis*, alpha-amylase produced by *Bacillus amyloliquefaciens* and polygalacturonase produced by *Aspergillus aculeatus*, and is intended for inclusion in feed rich in starch and non-starch polysaccharides (mainly arabinoxylans and beta-glucans). Avizyme 1500 was provided by Multi vita Animal Nutrition, Egypt.

Phyzyme XP-TPT10000 is 6-phytase (EC.3.1.3.26) contains 10000 U/g (supplied by Danisco Animal Nutrition, UK).

3.4. Measurements and methods of results' interpretation

3.4.1. Live weight gain (g)

Broilers were individually weighed, weekly to the nearest gram. Live body weight gain was calculated by subtracting the live body weight

in gram at the beginning of each experimental week from live body weight at the end the same experimental week.

3.4.2. Feed intake (g).

Average weekly feed intake was individually calculated by the difference between weekly offered feed and the weekly residual in g/replicate.

3.4.3. Feed conversion ratio (kg feed/kg gain)

It represents the mount of feed in kilograms required to produce one kilogram of weight gain.

3.4.4. Slaughter test

At the end of growth period (42 day of age), 6 birds were randomly taken from each treatment to carry out the slaughter test to determine the carcass traits. Before slaughtering, the birds were starved for 12 hours, then weighed, slaughtered and allowed to bleed freely for about 5 minutes. Absolute weight of carcass, heart, gizzard, liver, and abdominal fat were proportioned to the live body weight upon slaughtering.

3.4.5. Biochemical constituents of blood serum

Blood samples were collected from four birds of blood serum measurements. Concentrations of total protein (**Henry et al., 1974**), albumin (**Doumas et al., 1977**), total cholesterol (**Watson, 1960**), triglycerides (**Zollner and Kirsch, 1962**), glucose (**Trinder, 1969**), creatinine (**Husdan and Rapaport, 1968**), liver enzymes AST (aspartate amino transferase) and ALT (alanine amino transferase), according to

Retiman and Frankel (1957), were determined. Globulin was calculated by difference between total protein and albumin. Also, albumin to globulin ratio was calculated.

3.4.6. Production index

$$= \{[\text{daily weight gain} \times (100 - \text{mortality})]/(10 \times \text{feed conversion})\}.$$

3.4.7. Statistical procedures

Data from all response variables were subjected to a two factorial analysis using **SAS (1990)**. Variables having significant differences were compared using Duncan's Multiple Range Test (**Steel and Torrie, 1960**).

The following model was applied;

$$Y_{ij} = \mu + S_i + P_j + (SP)_{ij} + E_{ij}; \text{ where;}$$

Y_{ij} = Individual observation.

μ = Overall mean.

S_i = Effect of dietary ME level ($i = 1, 2$).

P_j = Effect of supplemental enzyme source ($j = 1, 2, 3, \text{ and } 4$).

$(SP)_{ij}$ = Effect of the interaction between dietary ME level.

and supplemental enzyme source.

E_{ij} = experimental error

4. RESULTS AND DISCUSSION

Data presented in Table (3) illustrate the main effect of dietary ME levels and supplemental enzyme source on growth performance (live weight gain and feed conversion ratio) of broilers during 0-3, 4-6, and 0-6 weeks of age. Initial live weights of broilers were not significantly affected by main factors under investigation, indicating the randomization of allocation process. Concerning, dietary ME level, it is clear that optimum dietary ME level had a superior on live growth performance rather than the low dietary ME level, in terms of significant ($p<0.01$) improved live weight gains and feed conversion ratios throughout the studied experimental intervals. Total weight gain and total feed conversion of optimum ME diet fed broilers were improved by 9.49% and 9.63 %, respectively over that of the low ME fed broilers. Enzyme supplementation had positive significant effect on growth performance of broilers, except for live weight gain during 0-3 weeks of age. It is worth noting that the combination between Avizyme 1500 + Phzyme XP TPT 10000 had inferior result on live weight gain during 0-3 weeks of age, it turned to the best formula during 4-6, and 0-6 weeks of age, while each enzyme source alone gave comparable results to the control concerning live weight gains. Regarding the effect of supplemental enzyme sources on feed conversion ratio, Phzyme XP TPT 10000 or Avizyme 1500 + Phzyme XP TPT 10000 significantly ($p<0.01$) were better than the control or Avizyme 1500 fed diet broilers either during 4-6 or 0-6 weeks intervals, while no significant effect during the 0-3 weeks had been reported for the studied enzyme sources.

Effect of the interaction between dietary ME level and supplemental enzyme source on live growth performance of broilers is presented in (4). Concerning the live weight gain, the broilers fed on optimum ME level within each treatment surpassed its equivalent group with low ME diet ($p < 0.01$) during the 0-3 weeks of age. The same was true during the 4-6 and 0-6 weeks of age period, except for the group fed low ME-Avizyme + Phyzyme which gave comparable results for its equivalent group on optimum ME level. Feed conversion ratio was not significantly affected by dietary treatments during 0-3 weeks of age, then turned in similar manner resemble that with weight gain results, where each treatment within the optimum ME level was better than its match within the low ME groups, except for the group fed low ME-Avizyme + Phyzyme that gave similar results to its equivalent on optimum ME diet.

Results recorded on the effect of dietary ME level on growth performance are in accord with those of **Cheng et al., (1997)**, **El-Husseini et al., (2002)**, **Maiorka et al., (2004)**, **Golian et al., (2010)** that live weight gain and feed conversion ratio of broilers were significantly improved as dietary ME level increased. **Downs et al., (2006)** reported no difference in live weight gain, but improved feed conversion ration with increasing dietary ME level of the broilers. **Jafarnejad and Sadegh (2011)** reported no effect on live weight gain of broilers with increasing the energy density of the diet. While, **El Tazi et al., (2009)** reported neither live weight gain nor feed conversion ratio were affected by dietary energy level.

Results on Avizyme 1500 agree with those reported by **Steenfeldt et al., (1998)** and **Lü et al., (2009)** that using xylanase enzyme supplementation had no effect on broiler performance. However, **Zanella et al., (1999)** reported that Avizyme[®]1500 supplemented to a maize-SBM based diet resulted in an increase of 1.9 % in live weight and improvement of 2.2 % in feed conversion ratio. **Pettersson and Aman (2007)** using an enzyme cocktail, consisting of xylanase and β -glucanase resulted in a significant increase in body-weight. **Owens et al., (2008)** compared between different xylanses sources and reported improved total live weight gain (+12%) and feed conversion ratio (+9%) for all enzyme-supplemented diets compared to the control fed broilers

It is clear that adding a phytase-enzyme source with more efficacious than adding a versatile of enzyme mixture (Avizyme 1500) to improve the growth performance of the broilers. Also, Phzyme XP TPT 10000 improved growth performance (feed conversion ratio) of broilers compared to the control. Nevertheless, the difference between live weight gains of broilers fed the diet supplemented with Phzyme XP TPT 10000 and the control did not reach significant level. Some researchers have shown significant increases in live body weight with phytase supplementation to broiler diets (**El-Saady, 1999, Zanini and Sazzad, 1999, Sohail and Roland, 1999, Cabahug et al., 1999, Raw et al., 1999, El-Deeb et al., 2000, Waldroup et al., 2000, Yan et al., 2000, Zyla et al., 2000, Abd El-Samee, 2002, El-Medany and El-Afifi, 2002, Balasubramanian et al., 2002, Lan et al., 2002, Qota et al., 2002, Salem et al., 2003, Abdo, 2004, Aggoor et al. 2004, El-Nagmy et al., 2004, Abou El-Wafa et al., 2005, Onyango et al., 2005, Watson et al., 2005, Pillai et al., 2006, Pirgozliev et al., 2007, Liu et al., 2008, Ravindran et al., 2008, Zhou et al., 2009, and**

Jalali et al., 2009), Others have demonstrated that phytase supplementation had no effect on live weight gain of broilers (**Vetési et al., 1998, Applegate et al. 2003, Miles et al., 2003, Karimi, 2005, and Akyurek et al., 2005**) or negatively affected growth performance (**Powell, 2005**).

Following the same manner, phytase supplementation significantly improved feed conversion ratio of broilers. These findings are in accordance with some other reports (**Vetési et al., 1998, El-Saady, 1999, Ravindran et al., 1999, Raw et al., 1999, Waldroup et al., 2000, Abd El-Samee, 2002, Balasubramanian et al., 2002, Lan et al., 2002, El-Nagmy et al., 2004, Aggoor et al., 2004, El-Nagmy et al., 2004, Hassanabadi et al., 2004, Liu et al., 2008, Ravindran et al., 2008, Zhou et al., 2009 and Jalali et al., 2009**). Other studies did not detect beneficial response to phytase supplementation on broilers' feed conversion ratio (**Sebastian et al., 1996, El-Deeb et al., 2000, Qota et al., 2002, Applegate et al., 2003, Salem et al., 2003, Karimi, 2005, and Akyurek et al., 2005**).

The improvement in feed conversion due to phytase supplementation may be interpreted on the base that phytate limited the availability of several nutrients such as minerals (**Farrel and Martin, 1998**), protein (**Cheryan, 1980**), energy (**Ravindran, 1999**) and digestive enzymes (**Deshpande and Cheryan, 1984**). The synergistic effect between Avizyme 1500 and Phzyme XP TPT 10000 is clear compared to the effect of each enzyme preparation, alone. It is assumed that the positive effect of supplementing Phzyme XP TPT 10000 (phytase enzyme) was further enhanced when combined with Avizyme 1500. The latter may be increased the digestibility of the nutrients, as it set free by the action of the phytase enzyme. The reduction in the amount of high-

molecular-weight arabinoxylan fragments leads to a decrease in gut viscosity and therefore an increase in the digestion and uptake of nutrients (**Andersson et al., 2003**).

Results provided in Table (5) indicate that dietary ME level had no significant effect on carcass characteristics of the broilers (carcass, liver, heart, gizzard, total edible parts, and abdominal fat percentages). Among studied supplemental enzyme sources, Phyzyme XP TPT 10000, alone significantly increased carcass percentage compared to the control ($p<0.05$). The latter, had significantly ($p<0.01$) the highest liver, and gizzard percentages. Each enzyme source, separately, showed lower abdominal fat percentage ($p<0.05$) compared to the control or enzyme combination source. Heart or total edible part percentage did not significantly affected by enzyme source.

Results presented in Table (6) indicate that the interaction between dietary ME level and supplemental enzyme source had no significant effect on any of the studied carcass traits (carcass, liver, heart, gizzard, total edible parts and abdominal fat percentages).

In agreement with the current findings, **Maiorka et al., (2004)** reported that dietary ME level (3200 vs.2900 kcal/kg) had no significant effect on carcass percentage of broilers. While, abdominal fat percentage was decreased with the high dietary ME level. **Downs et al., (2006)** found that carcass yield and abdominal fat percentages were not significantly affected by dietary ME level. Also, **El Tazi et al., (2009)** agree that dietary energy level had no significant effect on dressing percentage of broilers.

Little studies focused on the effect of supplemental enzyme mixture on carcass traits. In agreement with the results of the current study **Saleh et al., (2005)** found that the addition of cellulase led to a significant decrease in the abdominal fat pad weight of the broilers. **Zanella et al. (1999)** also looked at the effect of enzyme supplementation (xylanase, amylase, and protease) on the abdominal fat content of broilers, no significant results were found.

Concerning the effect on carcass characteristics, the supplemental enzyme sources had no effect on total edible parts percentage. In agreement with the present findings, **El-Saady (1999)**, **Scheideler and Ferket (2000)**, **El-Medany and El-Afifi (2002)**, **Abdo (2004)**, **El-Nagmy et al., (2004)** found that phytase did not significantly affect carcass yield. While, **Salem et al., (2003)**, and **Pillai et al., (2006)** found that phytase supplementation significantly increased dressed carcass percentage and abdominal fat of broilers compared with those fed the control diet without phytase supplementation. **Viveros et al., (2002)** reported that the inclusion of phytase in diets had no significant effect on dressing percentage of the broilers. However, **Scheideler and Ferket (2002)**, and **El-Medany et al., (2002)** stated that the addition of phytase to diet improved significantly the dressing percentage of the broilers.

Results on the effect of dietary ME level or supplemental enzyme sources on blood serum constituents (total protein, albumin, globulin, albumin/globulin ratio, glucose, triglycerides, cholesterol, creatinine, ALT and AST) are shown in Table (7). Optimum ME level significantly increased serum total protein ($p<0.05$), globulin ($p<0.01$), glucose ($p<0.01$), triglycerides ($p<0.05$), ALT ($p<0.01$), and AST ($p<0.05$), while

decreased albumin/globulin ratio ($p < 0.01$) compared to low ME level. On the other hand, supplemental enzyme level had no significant effect on all studied serum parameters, except the kidneys' creatinine ($p < 0.05$) as the control fed broilers showed the highest level.

Results on the effect of the interaction between dietary ME level and supplemental enzyme source on serum constituents of broilers are provided in Table (8). Dietary treatments significantly affected serum total protein ($p < 0.05$), globulin ($p < 0.05$), glucose ($p < 0.01$), and creatinine ($p < 0.05$). Serum albumin, albumin/globulin ratio, triglycerides, cholesterol, ALT and AST were not significantly affected by dietary treatments.

No data are available considering the effect of dietary energy level on the studied blood constituents. Phytase had no effect on plasma, total protein, total lipids, and cholesterol, which agree with other reports (**Kornegay et al., 1998**, **Attia et al., 2001**, and **Qota et al., 2002**). However, **Jalali et al., (2009)** found that serum total protein and albumin concentrations were increased by phytase addition. Concerning liver functions, **Viveros et al., (2002)** reported that phytase supplementation increased serum AST but decreased serum ALT activity. Meanwhile, **Abdo (2004)** found that phytase in broiler diet had no significant effect on AST activity. **Selim (2007)** found that phytase decreased plasma cholesterol of broilers.

Results presented in Table (9) reveal the economic index study. It is clear that feeding broilers on optimum dietary energy level was better (331.4) than feeding the low energy diet (275.0). Concerning supplemental enzyme sources, feeding a phytase enzyme alone or combined with enzymes' mixture was rather than feeding the enzymes'

mixture alone or feeding diets without supplemental enzymes in leveling up the performance index. Individual treatments had fluctuated effect on economic study. Alongside the optimum energy level, all supplemental enzyme sources showed higher economic index than the control group without enzyme supplementation. Only feeding a phytase enzyme combined with the enzymes' mixture showed the highest economic index, while other supplemental enzymes' source failed to increase the economic index over that obtained by the control with low energy level.

Table (3): Main effect of dietary energy levels and enzyme sources on live weight gain and feed conversion ratio of broilers

		ME level		Significance	Enzyme source				Significance
		Optimum	Low		Control	Avizyme 1500	Phyzyme XP TPT 10000	Avizyme 1500 +Phyzyme XP TPT 10000	
Initial live weight		40±0.08	40±0.07	ns	40±0.09	40±0.09	40±0.14	40±0.11	ns
Live weight gain	0-3 wks	833 ^A ±5.45	779 ^B ±5.95	**	810 ^A ±15	809 ^A ±16	817 ^A ±5.56	787 ^B ±10	*
	4-6 wks	1532 ^A ±17	1381 ^B ±28	**	1425 ^C ±26	1400 ^C ±39	1467 ^B ±63	1533 ^A ±14	**
	0-6 wks	2365 ^A ±18	2160 ^B ±27	**	2236 ^{BC} ±41	2209 ^C ±54	2285 ^{AB} ±67	2320 ^A ±17	**
Feed conversion ratio	0-3 wks	1.37 ^B ±0.01	1.47 ^A ±0.01	**	1.46±0.02	1.40±0.02	1.40±0.02	1.43±0.03	ns
	4-6 wks	2.01 ^B ±0.02	2.27 ^A ±0.04	**	2.22 ^A ±0.03	2.22 ^A ±0.07	2.16 ^B ±0.10	1.97 ^C ±0.02	**
	0-6 wks	1.69 ^B ±0.01	1.87 ^A ±0.02	**	1.84 ^A ±0.02	1.81 ^{AB} ±0.05	1.78 ^B ±0.05	1.70 ^C ±0.02	**

^{A-C} Means within the same row differ significantly, ns=not significant, *= $P>0.05$, **= $P>0.01$

Table (4): Effect of dietary treatments on live weight gain and feed conversion ratio of broilers

	Initial live weight	Live body weight gain			Feed conversion ratio		
		0-3 wks	4-6 wks	0-6 wks	0-3 wks	4-6 wks	0-6 wks
Optimum ME-Control	40 ±0.11	847 ^a 11±	1485 ^c 18±	2333 ^b 29±	1.41 0.01±	2.16 ^c 0.04±	1.78 ^b 0.02±
Optimum ME-Avizyme1500 [®]	40 ±0.15	849 ^a 5.5±	1500 ^c 17±	2349 ^b 22±	1.33 0.01±	2.02 ^d 0.01±	1.68 ^{cd} 0.01±
Optimum ME-Phyzyme XP TPT 10000	40.42 ±0.22	827 ^{ab} 5.42±	1633 ^a 19±	2460 ^a 24±	1.36 0.02±	1.88 ^e 0.01±	1.62 ^d 0.00±
Optimum ME-Avizyme1500+Phyzyme XP TPT10000	40.35 ±0.18	809 ^b 7.00±	1509 ^{bc} 15±	2318 ^b 22±	1.37 0.04±	1.99 ^d 0.04±	1.68 ^{cd} 0.04±
Low ME-Control	40.25 ±0.15	774 ^c 5.5±	1366 ^d 25±	2140 ^c 30±	1.51 0.01±	2.27 ^b 0.04±	1.89 ^a 0.02±
Low ME-Avizyme1500 [®]	40.12 ±0.11	769 ^c 11.9±	1300 ^e 13.5±	2070 ^c 21.9±	1.47 0.01±	2.41 ^a 0.01±	1.94 ^a 0±
Low ME-Phyzyme XP TPT 10000	40.30 ±0.20	807 ^b 7.13±	1301 ^e 14.5±	2109 ^c 14.6±	1.43 0.04±	2.43 ^a 0.01±	1.93 ^a 0.01±
Low ME-Avizyme1500+Phyzyme XPTPT10000	40.30 ±0.15	766 ^c 10.9±	1556 ^b 19.19±	2322 ^b 29.5±	1.48 0.04±	1.96 ^{de} 0.01±	1.72 ^{bc} 0.02±
Significance	ns	**	**	**	ns	**	**

^{a-c} Means within the same column differ significantly, ns=not significant, **= $P > 0.01$.

Table (5): Main effect of dietary energy levels and enzyme sources on carcass characteristics

Item	ME level		Sig.	Enzyme source				Sig.
	Optimum	Low		Without	Avizyme 1500	Phyzyme XP TPT 10000	Avizyme 1500+Phyzyme XP TPT 10000	
Carcass %	73.3±0.3	72.7±0.2	ns	72.6 ^B ±0.4	73.2 ^{AB} ±0.5	73.7 ^A ±0.3	72.4 ^B ±0.4	*
Liver %	1.98±0.04	1.89±0.04	ns	2.12 ^A ±0.05	1.94 ^B ±0.05	1.76 ^C ±0.04	1.94 ^B ±0.06	**
Heart %	0.54±0.02	0.54±0.02	ns	0.59±0.02	0.52±0.02	0.52±0.04	0.53±0.03	ns
Gizzard %	1.41±0.05	1.48±0.03	ns	1.57 ^A ±0.06	1.35 ^B ±0.06	1.39 ^B ±0.05	1.47 ^{AB} ±0.07	*
Total edible part %	77.2±0.3	76.6±0.2	ns	76.9±0.3	77.1±0.4	77.4±0.3	76.3±0.3	ns
Abdominal fat %	2.11±0.05	2.21±0.05	ns	2.17 ^{AB} ±0.09	2.04 ^B ±0.05	2.10 ^B ±0.06	2.33 ^A ±0.08	*

^{A-C} Means within the same row differ significantly, ns=not significant, *= $P>0.05$, **= $P>0.01$

Table (6): Effect of dietary treatments on carcass characteristics

Treatment	Carcass %	Liver %	Heart %	Gizzard %	Total edible %	Abdominal fat %
Optimum ME-Control	72.6 ±0.06	2.09 ±0.08	0.60 ±0.02	1.62 ±0.11	77.00 ±0.54	2.16 ±0.15
Optimum ME-Avizyme 1500 [®]	73.32 ±0.87	2.02 ±0.05	0.53 ±0.03	1.31 ±0.08	77.20 ±0.84	2.03 ±0.07
Optimum ME-Phyzyme XP TPT 10000	74.20 ±0.58	1.77 ±0.06	0.52 ±0.05	1.36 ±0.09	77.87 ±0.44	2.05 ±0.09
Optimum ME-Avizyme1500+Phyzyme XP TPT 10000	73.05 ±0.39	2.01 ±0.09	0.50 ±0.02	1.33 ±0.06	76.91 ±0.35	2.20 ±0.07
Low ME-Control	72.48 ±0.43	2.13 ±0.06	0.58 ±0.04	1.52 ±0.04	76.73 ±0.46	2.16 ±0.09
Low ME-Avizyme 1500 [®]	73.16 ±0.36	1.85 ±0.08	0.49 ±0.02	1.38 ±0.09	76.90 ±0.35	2.04 ±0.05
Low ME-Phyzyme XP TPT 10000	73.25 ±0.13	1.73 ±0.05	0.51 ±0.05	1.41 ±0.05	76.91 ±0.11	2.15 ±0.06
Low ME-Avizyme1500+Phyzyme XP TPT10000	71.70 ±0.46	1.85 ±0.06	0.56 ±0.04	1.59 ±0.09	75.72 ±0.38	2.46 ±0.11
Significance	Ns					

ns = not significant

Table (7): Main effect of dietary energy levels and enzyme sources on some blood serum constituents

Constituent	ME level		Significance	Enzyme source				Significance
	Optimum	Low		Control	Avizyme 1500	Phyzyme XP TPT 10000	Avizyme 1500+Phyzyme XP TPT 10000	
Total Protein	5.45 ^A ±0.14	4.94 ^B ±0.14	*	4.69±0.23	5.25±0.20	5.33±0.19	5.23±0.26	ns
Albumin (A)	3.08±0.05	3.00±0.09	ns	2.96±0.12	3.06±0.09	3.02±0.07	3.11±0.14	ns
Globulin (G)	2.38 ^A ±0.09	1.99 ^B ±0.07	**	2.00±0.12	2.26±0.14	2.31±0.13	2.17±0.13	ns
A/G ratio	1.30 ^B ±0.04	1.48 ^A ±0.03	**	1.46±0.06	1.34±0.14	1.31±0.05	1.45±0.05	ns
Glucose	127 ^A ±5.93	104 ^B ±5.04	**	109±7.55	122±9.05	122±9.31	199±9.47	ns
Triglycerides	88 ^A ±1.20	85 ^B ±0.87	*	86±1.30	89±1.82	86±1.04	86±1.92	ns
Cholesterol	74±2.74	79±2.16	ns	79±3.39	77±4.20	71±2.74	81±3.32	ns
Creatinine	0.22±0.01	0.18±0.01	ns	0.25 ^A ±0.02	0.17 ^B ±0.01	0.20 ^{AB} ±0.02	0.18 ^B ±0.01	*
ALT	2.71 ^A ±0.18	1.92 ^B ±0.16	**	2.40±0.24	2.66±0.33	2.08±0.15	2.12±0.36	ns
AST	235 ^A ±17	174 ^B ±17	*	217±25	233±24	196±22	172±33	ns

^{A-B} Means within the same row differ significantly, ns=not significant, *= $P>0.05$, **= $P>0.01$

Table (8): Effect of dietary treatments on some blood serum constituents

Treatment	Total Protein	Albumin (A)	Globulin (G)	A/G ratio	Glucose	Triglycerides	Cholesterol	Creatinine	ALT	AST
Optimum ME-Control	5.37 ^{abc} ±0.24	3.15 ±0.08	2.22 ^{abc} ±0.17	1.36 ±0.08	100 ^b ±6.48	86 ±2.39	73 ±5.39	0.22 ^{abc} ±0.03	2.47 ±0.39	207 ±43
Optimum ME-Avizyme1500 [®]	5.72 ^a ±0.17	3.12 ±0.04	2.60 ^a ±0.14	1.21 ±0.06	143 ^a ±5.54	92 ±1.49	75 ±7.08	0.19 ^{bc} ±0.02	3.17 ±0.30	265 ±30
Optimum ME-Phyzyme XP TPT 10000	5.72 ^a ±0.13	3.12 ±0.04	2.60 ^a ±0.09	1.20 ±0.03	122 ^{ab} ±13.00	86 ±1.79	69 ±4.73	0.25 ^{ab} ±0.03	2.42 ±0.17	217 ±38
Optimum ME-Avizyme1500+Phyzyme XP TPT10000	5.00 ^{abc} ±0.40	2.92 ±0.20	2.10 ^{bc} ±0.20	1.42 ±0.08	142 ^a ±7.00	89 ±3.09	80 ±5.12	0.22 ^{abc} ±0.01	2.77 ±0.54	252 ±29
Low ME-Control	4.55 ^c ±0.28	2.77 ±0.19	1.77 ^c ±0.11	1.56 ±0.06	119 ^{ab} ±12.80	87 ±1.43	85 ±1.65	0.28 ^a ±0.04	2.32 ±0.34	228 ±31
Low ME-Avizyme1500 [®]	4.77 ^{bc} ±0.14	3.00 ±0.18	1.92 ^{bc} ±0.04	1.4 ±0.07	101 ^b ±7.36	86 ±2.59	79 ±5.43	0.16 ^{bc} ±0.02	2.15 ±0.50	202 ±36
Low ME-Phyzyme XP TPT 10000	4.95 ^{abc} ±0.25	2.92 ±0.13	2.02 ^{bc} ±0.13	1.41 ±0.06	102 ^b ±12.92	85 ±1.31	73 ±3.17	0.15 ^c ±0.03	1.75 ±0.10	174 ±22
Low ME-Avizyme1500+Phyzyme XP TPT 10000	5.5 ^{ab} ±0.34	3.30 ±0.16	2.25 ^{ab} ±0.18	1.48 ±0.05	96 ^b ±4.95	83 ±1.08	82 ±5.01	0.13 ^c ±0.01	1.47 ±0.17	92 ±3.49
Significance	*	ns	*	ns	**	ns	ns	*	ns	ns

^{a-c} Means within the same column differ significantly, ns=not significant, *= $P>0.05$, **= $P>0.01$

Table (9): Economic study

Main effect	Performance index	Treatment	Performance index
Energy level		Optimum ME-Control	312.1
Optimum ME	331.4	Optimum ME-Avizyme1500®	332.9
Low ME	275.0	Optimum ME-Phyzyme XP TPT 10000	361.6
Enzyme source		Optimum ME-Avizyme1500+Phyzyme XP TPT10000	328.5
Without	289.3	Low ME-Control	269.6
Avizyme1500®	285.6	Low ME-Avizyme1500®	254.1
Phyzyme XP TPT 10000	305.6	Low ME-Phyzyme XP TPT 10000	260.2
Avizyme1500+Phyzyme XP TPT 10000	324.9	Low ME-Avizyme1500+Phyzyme XP TPT 10000	321.4

4. SUMMARY

The work aimed at comparative study to investigate different enzyme sources on the energy utilization in broilers fed wheat-base diets.

Three hundred and twenty one-day-old unsexed Cobb chicks were randomly distributed between 8 experimental groups of 40 chicks, each to study to investigate the effect of supplementing different enzyme sources (phytase *vs.* enzyme mixture) to different dietary metabolisable energy levels (optimum *vs.* low) by broilers fed wheat-based diets on growth performance, carcass traits and some blood serum constituents. Diets were as follows:

Treat. 1: Diet containing the ideal ME energy level without enzyme supplement (control).

Treat. 2: Control + Avizyme 1500 (100 g/100 kg)

Treat. 3: Control + Phyzyme XP TPT 10000 (50g /100 kg)

Treat.4: Control + Avizyme 1500 (50 g) + Phyzyme XP TPT 10000 (25 g /100 kg)

Treat. 5: Diet containing a low-ME (low 100 kcal/kg diet) without enzyme supplement.

Treat. 6: Low-ME diet + Avizyme 1500 (100 g / 100 kg)

Treat.7: Low-ME diet + Phyzyme XP TPT 10000 (50g /100 kg)

Treat. 8: Low-ME diet + Avizyme 1500 (50 g) + Phyzyme XP TPT 10000 (25g / 100 kg).

Results could be summarized as follows:

- Optimum dietary ME level had a superior on live growth performance rather than the low dietary ME level, in terms of significant ($p<0.01$)

improved live weight gains and feed conversion ratios throughout the studied experimental intervals.

-Enzyme supplementation had positive significant effect on growth performance of broilers, except for live weight gain during 0-3 weeks of age.

- Dietary ME level had no significant effect on carcass characteristics of the broilers.

- Among studied supplemental enzyme sources, Phyzyme XP TPT 10000, alone significantly increased carcass percentage compared to the control ($p < 0.05$). The latter, had significantly ($p < 0.01$) the highest liver, and gizzard percentages. Each enzyme source, separately, showed lower abdominal fat percentage ($p < 0.05$) compared to the control or enzyme combination source. Heart or total edible part percentage did not significantly affected by enzyme source.

- Optimum ME level significantly increased serum total protein ($p < 0.05$), globulin ($p < 0.01$), glucose ($p < 0.01$), triglycerides ($p < 0.05$), ALT ($p < 0.01$), and AST ($p < 0.05$), while decreased albumin/globulin ratio ($p < 0.01$) compared to low ME level.

-Supplemental enzyme level had no significant effect on all studied serum parameters, except the kidneys' creatinine ($p < 0.05$) as the control fed broilers showed the highest level.

- Feeding optimum energy level diet had higher economical index than feeding low energy diet.

-Feeding a phytase enzyme alone or combined with enzymes' mixture give higher economical index than feeding the enzymes' mixture alone or feeding diets without supplemental enzymes

5. REFERENCES

- Abd El-Samee, M. (2002).** Effect of different levels of crude protein, sulphur amino acids, microbial phytase and their interaction on broiler chick performance. *Egypt. Poult. Sci.*, 22: 999-1021.
- Abdo, Zeinab (2004).** Effect of phytase supplementation on the utilization of nigella sativa seed meal in broiler diets. *Egypt. Poult. Sci.*, 24:143-162.
- Abou El-Wafa, S., El-Husseiny, O., and Shabaan, M. (2005).** Influence of microbial phytase and energy levels on broiler performance fed low-phosphorus diets. 3rd Int. Poultry Conference 4-7 Apr., Egypt.
- Acamovic, T. (2001).** Commercial application of enzyme technology for
(2003). Effect of endoxylanase-containing enzyme preparations and laccase on the solubility of rye bran arabinoxylan. *J. Sci. Food Agric.*, 83(7): 617-623.
- Aggoor, F., Attia, Y., Qota, E., and Abd El-Moaty, H. (2004).** Effect of enzyme supplementation and dietary formulation on digestible amino acid on performance; meat quality and plasma constituents of roaster type chicks fed. 1: High level of rapeseed meal. World's Poultry Congress, 8-13 June, Istanbul, Turkey.
- Akyurek, H., Senkoylu, N., and Ozduven, M. (2005).** Effect of microbial phytase on growth performance and nutrients digestibility in broilers. *Pakistan J. of Nutrition*, 4:22-26.

Andersson, R., Eliasson, C., Selenare, M., Kamal-Eldin, A., and Åman, P. (2003). Effect of endoxylanase-containing enzyme preparations and laccase on the solubility of rye bran arabinoxylan. *J. Sci. Food Agric.* 83(7): 617-623.

Applegate, T., Joern, B., Nussbaum-Wagler, D., and Angel, R. (2003).

Water-soluble phosphorus in fresh litter is dependent upon phosphorus concentration fed but not on fungal phytase supplementation. *Poult. Sci.*, 82:1024-1029.

Attia, Y., Abd El-Rahman, S., and Qota, E. (2001). Effects of microbial phytase with or without cell-wall splitting enzymes on the performance of broilers fed suboptimum levels of dietary protein and metabolizable energy. *Egypt. Poult. Sci.* 21, 521-547.

Balasubramanian, D., Jalaludee, A., Peethambaran, P., and Amritha

Viswanath (2002). Influence of microbial phytase on growth performance in broiler chicken. *Indian J. Anim. Sci.*; 72: 1048-1050.

Bedford, M. R. (1993). Mode of action of feed enzymes. *The Journal of Appl. Poult. Res.*, 2(1):85.

Bedford, M. R. (2000). Exogenous enzymes in monogastric nutrition—their current value and future benefits. *Anim. Feed Sci. and Tech.*, 86: 1-13.

- Bedford, M.R., and Classen, H. L. (1992).** Reduction of intestinal viscosity through manipulation of dietary rye and pentosanase concentrations is effected through changes in the carbohydrate composition of the intestinal aqueous phase and results in improved growth rate and food conversion efficiency of broiler chicks. *J. Nutr.*, 122: 560-569
- Bedford, M. R., and Schulze H. (1998).** Exogenous enzymes for pigs and poultry. *Nutrition Research Reviews*, 11(01): 91-114.
- Beg, Q., Kapoor, M., Mahaja, L., and Hoondal, G. (2001).** Microbial xylanases and their industrial applications: A review. *Appl. Microbiol. Biotechnol.*, 56(3): 326-338.
- Bhat, M. (2000).** Cellulases and related enzymes in biotechnology. *Biotechnol. Adv.*, 18(5): 355-383.
- Boros, D., Marquardt, R., and Guenter, W. (1998).** Site of exoenzyme action in gastrointestinal tract of broiler chicks. *Canadian J. Anim. Sci.*, 78: 599-602.
- Botha, C. (2011).** The use of fibrolytic enzymes in maize-soya based broiler diets. M.Sc. Thesis, Stellenbosch University, South Africa.
- Brufau, J., Francesch, M., and Pérez-Vendrel, A. (2001).** Are we making the best use of NSP enzymes? *Feed Mix*, 9(6): 37.
- Cabahug, S., Ravindran, V., Selle, P., and Bryden, W. (1999).** Response of broiler chickens to microbial phytase supplementation as influenced by dietary phytic acid and non phytate phosphorous content, 1- Effect on bird performance and toe ash. *Brit. Poult. Sci.*, 40: 660-666.

- Carre, B., Gomez, J., and Chagneau, M. (1995).** Contribution of oligosaccharide and polysaccharide digestion, and excreta losses of lactic acid and short chain fatty acids, to dietary metabolisable energy values in broiler chickens and adult cockerels. *Brit. Poult. Sci.*, 36:611-629.
- Castanon, J., Flores, M., and Pettersson, D. (1997).** Mode of degradation of non-starch polysaccharides by feed enzyme preparations. *Anim. Feed Sci. Tech.*, 68(3-4): 361-365.
- Cheng, T., Hamre M., and Coon C. (1997).** Effect of environmental temperature, dietary protein, and energy levels on broiler performance. *J. Appl. Poultry Res.*, 6:1-17.
- Cheryan, M. (1980).** Phytic acid interactions in food systems. *CRC. Critical Reviews in Food and Nutrition*, 13: 297-335.
- Chesson, A. (2001).** Non-starch polysaccharide degrading enzymes in poultry diets: Influence of ingredients on the selection of activities. *Worlds' Poult. Sci. J.*, 57(3): 251-263.
- Choct, M. (2006).** Enzymes for the feed industry: Past, present and future. *Worlds' Poult. Sci. J.* 62(1): 5-16.
- Choct, M., Hughes R., and Bedford M. (1999)** Effects of xylanase on individual bird variation, starch digestion throughout the intestine and ileal and caecal volatile fatty acid production in chickens fed wheat. *Brit. Poult. Sci.*, 40: 419–422.
- Choct, M., Hughes, R., Trimble, R., Angkanaporn, K., and Annison G. (1995).** Non-starch polysaccharide-degrading enzymes increase the performance of broiler chickens fed wheat of low apparent metabolizable energy. *J. Nutr.*, 125(3): 485.

- Choct, M., Hughes R., Wang, J., Bedford, R., Morgan, J., and Annison, G. (1996).** Increased small intestinal fermentation is partly responsible for the anti-nutritive activity of non-starch polysaccharides in chickens. *Brit. Poult. Sci.* 37(3): 609-621.
- Classen, H. (1996).** Cereal grain starch and exogenous enzymes in poultry diets. *Anim. Feed Sci. Tech.*, 62(1): 21-27.
- Cowieson, A. J. (2005).** Factors that affect the nutritional value of maize for broilers. *Anim. Feed Sci. Tech.*, 119:293–305.
- Cowieson, A., and Ravindran, V. (2008).** Effect of exogenous enzymes in maize-based diets varying in nutrient density for young broilers: Growth performance and digestibility of energy, minerals and amino acids. *Brit. Poult. Sci.*, 49:37–44.
- Cowieson, A., Hruby M., and Isaksen, M. (2005).** The effect of conditioning temperature and exogenous xylanase addition on the viscosity of wheat-based diets and the performance of broiler chickens. *Brit. Poult. Sci.*, 46(6): 717-724.
- Cowieson, A. J., Singh D., and Adeola, O. (2006).** Prediction of ingredient quality and the effect of a combination of xylanase, amylase, protease and phytase in the diets of broiler chicks. 1. Growth performance and digestible nutrient intake. *Brit. Poult. Sci.*, 47:477-489.
- Deshpande, S., and Cheryan, M. (1984).** Effects of phytic acid, divalent cations and their interactions on alpha-amylase activity. *J. Food Sci.*, 49:516-519.

- Doumas, B. T., Watson, W. A., and Biggs, H. G. (1977).** Albumin standards and the measurement of serum albumin with bromocresol green. *Clin. Chem. Acta.*, 31: 87-96.
- Downs, K., Lien, R., Hess, J., Bilgili, S., and Dozier, W. (2006).** The effects of photoperiod length, light intensity, and feed energy on growth responses and meat yield of broilers. *J. Appl. Poult. Res.* 15:406-416.
- Eardman, J. (1979).** Oilseed phytates: Nutritional implications. *J. Amer. Oil Chem. Soc.*, 56: 736–741.
- Edwards, H. (1991).** Effects of phytase utilization by monogastric animals. *Proc. Georgia Nutrition Conference for Feed Manufactures.* Atlanta, pp. 1-6.
- El-Deeb, Mariam, Sharara, H., and Makled, M. (2000).** Enhance calcium and phosphorus utilization by enzyme phytase supplemented to broiler diet contained rice bran. *Egypt. Poult. Sci.*, 20: 545-566.
- El-Husseini O., Shalash S., and Azouz H. (2002).** Response of broiler performance to diets containing hot pepper, and or fenugreek at different metabolizable energy levels. *Egypt. Poult. Sci.*, 22: 387-406.
- El-Medany, N., and El-Afifi, S. (2002).** The effect of microbial phytase on utilization of protein and phosphorus in broiler rations. *Egypt. Poult. Sci.*, 22: 427-443.
- El-Nagmy, K., Abd El-Samee, M., and Ibrahim, M. (2004).** Effect of dietary plant protein and microbial phytase levels on performance of broiler chicks. *Egypt. Poult. Sci.*, 24:101-121.

- El-Saady, M. (1999)** Effect of using local feedstuffs and phytase supplementation on the performance of broiler chicks. Egypt. Poult. Sci., 19: 939-954.
- El Tazi, S., Mohamed, K., and Mohamoud, A. (2009).** The effect of dietary microbial phytase and metabolizable energy levels on the performance of broiler chicks. J. Appl. Sci. Res., 5(12): 2096-2101.
- Engberg, R., Hedemann, M., Steinfeldt, S., and Jensen, B. (2004).** Influence of whole wheat and xylanase on broiler performance and microbial composition and activity in the digestive tract. Poult. Sci., 83: 925-938.
- Evers, A.D., Blakeney, A.B., and O'Brien, L.O. (1999).** Cereal structure and composition. Australian J. Agric. Res., 50:629-650.
- Farrel, D., and Martin, E. (1998).** Strategies to improve the nutritive value of rice bran in poultry diets. 3. The addition of inorganic phosphorus and phytase to duck diets. Brit. Poult. Sci., 39:601-611.
- Fourie, J. L. (2007).** The effects of a multiple-enzyme combination in maize-soya diets for broiler chickens. M.Sc. Thesis, Univ. Stellenbosch, South Africa.
- Francesch, M., and Geraert, P. (2009).** Enzyme complex containing carbohydrases and phytase improves growth performance and bone mineralization of broilers fed reduced nutrient corn-soybean-based diets. Poult. Sci., 88: 1915–1924
- Fuente, J., Perez de Ayala P., and Villamide M. (1995).** Effect of dietary enzyme on the metabolizable energy of diets with increasing levels of barley fed to broilers at different ages. Anim. Feed Sci. and Techno., 56:45–53.

- Godoy, S., Chicco, C., Meschy, F., and Fanny, R. (2005).** Phytic phosphorus and phytase activity of animal Feed Ingredients. INCI 30: 24-28.
- Golian, A., Azghadi, M., and Pilevar, M. (2010).** Influence of various levels of energy and protein on performance and humoral immune responses in broiler chicks. *Global Veterinaria*, 4 (5): 434-440.
- Greiner, R., and Konietzny, U. (2006).** Phytase for food application. *Food Technol., and Biotechnol.*, 44 (2):125-140.
- Gutierrez, A., Verstegen, M., den Hartog, L., Perez de Ayala, P., and Villamide, M. (2008).** Effect of wheat cultivar and enzyme addition to broiler chicken diets on nutrient digestibility, performance, and apparent metabolizable energy content. *Poult. Sci.*, 87:759–767
- Hassanabadi, A., Moghaddam, H., and Pourreza, J. (2004).** Effect of microbial phytase on apparent digestibility of amino acids and performance of male broiler chickens. *Agric. Sci. and Tech.*, 18: 49-56.
- Henry, R. J. (1974).** In *Clinical Chemistry. Principles and Technics.* Harper and Row edition, Maryland, USA.
- Hetland, H., Choct, M., and Svihus, B. (2004).** Role of insoluble non-starch polysaccharides in poultry nutrition. *Worlds' Poult. Sci. J.*, 60(4): 415-422.
- Husdan, H., and Rapaport A. (1968).** Estimation of creatinine by the Jaffa reaction. *Clin. Chem.*, 14:222-228.

- Huyghebaert, G. (1997).** The effect of a wheat-fat interaction on the efficacy of a multienzyme preparation in broiler chickens. *Anim. Feed Sci. and Technol.*, 68:55-66.
- Jafarnejad, S., and Sadegh, M. (2011).** The effects of different levels of dietary protein, energy and using fat on the performance of broiler chicks at the end of the third weeks. *Asian J. Poult. Sci.*, 5 (1): 35-40.
- Jalali, S., Jafarian, M., and Tavakoli, S. (2009).** Effects of supplemental dietary phytase on performance and some blood biochemical parameter of broiler chicks. 5th Int. Poult. Conf., Egypt, pp. 675-679.
- Karimi, A. (2005).** Effect of different non-phytate phosphorus levels and phytase sources on performance in broiler chicks. *Int. J. Poult. Sci.*, 4 (12):1001-1005.
- Kasim, A., and Edwards, H. (1998).** The analysis for inositol phosphate forms in feed ingredients. *J. Sci. Food Agric.* 76:1–9.
- Kocher, A., Choct, M., Ross, G., Broz, J., and Chung, T. (2003).** Effect of enzyme combinations on apparent metabolizable energy of corn-soybean meal-based diets in broilers. *J. Appl. Poult. Res.*, 12: 275 – 283.
- Kornegay, E., Denbow, D., and Ravindran, V. (1996).** Responses of broilers to graded levels of Natuphos phytase added to corn-soybean meal based diets containing three levels of non-phytate phosphorus. *Brit. J. Nut.* 75: 839-852.

- Kornegay, E., Zhang, Z., and Denbow, D. (1998).** Influence of microbial phytase supplementation of a low protein/amino acid diet on performance, ileal digestibility of protein and amino acids, and carcass measurements of finishing broilers, in: M. Coelho and Kornegay, E. Eds., *Phytase in Animal Nutrition and Waste Management*, 2nd Ed., BASF Cooperation, Mount Olive, NJ., USA.
- Knudsen, K. E. (1997).** Carbohydrate and lignin contents of plant materials used in animal feeding. *Anim. Feed Sci. Technol.* 67(4): 319-338.
- Kwakernaaka, C., van der Klis, J., and Brozb, J. (2011).** Effect of the addition of an endo-xylanase in wheat based diets on production performance of turkeys. *Proc. 18th European Symposium Poultry Nutrition*, Izmir, Turkey, pp. 80-81.
- Lan, G., Abdullah, N., Jalaludin, S., and Ho, Y. (2002).** Efficacy of supplementation of a phytase-producing bacterial culture on the performance and nutrient use of broiler chickens fed corn-soybean meal diet. *Poult. Sci.*, 81:1522-1532.
- Liu, N., Ru, Y., Cowieson, A., Li, F., and Cheng, H. (2008).** Effects of phytate and phytase on the performance and immune function of broilers fed nutritionally marginal diets. *Poult. Sci.*, 87:1105-1111.
- Lott, J., Ockenden, A., Raboy, V., and Batten, G. (2000).** Phytic acid and phosphorus in crop seeds and fruits: A global estimate. *Seed Sci. Res.*, 10:11–33.
- Lü, M., Li, D., Gong, L., Ru, Y., and Ravindran, V. (2009).** Effects of supplemental microbial phytase and xylanase on the performance of broilers fed diets based on corn and wheat. *Japanese Poult. Sci.*, 46: 217-223.

- Maiorka, A., Dahlke, F., Santin, E., Kessler, A., and Penz, Jr. (2004).** Effect of energy levels of diets formulated on total or digestible amino acid basis on broiler performance. *Brazilian J. Poult. Sci.*, 6 (2): 87- 91.
- Marron, L., Bedford, M., and McCracken, K. (2001).** The effects of adding xylanase, vitamin C and copper sulphate to wheat-based diets on broiler performance. *Brit. Poult. Sci.* 42(4): 493-500.
- Marsman, G., Gruppen, H., Van der Poel, A., Kwakkel, R., Verstegen, M., and Voragen, A. (1997).** The effect of thermal processing and enzyme treatments of soybean meal on growth performance, ileal nutrient digestibilities, and chyme characteristics in broiler chicks. *Poult. Sci.* 76(6): 864.
- Meng, X., Slominski, B., and Guenter, W. (2004).** The effect of fat type, carbohydrate, and lipase addition on growth performance and nutrient utilization of young broilers fed wheat-based diets. *Poult. Sci.*, 83: 1718-1727
- Meng, X., Slominski, B., Nyachoti, C., Campbell, L., and Guenter, W. (2005).** Degradation of cell wall polysaccharides by combinations of carbohydrase enzymes and their effect on nutrient utilization and broiler chicken performance. *Poult. Sci.*, 84: 37-47.
- Miles, D., Moore, P., Smith, Jr., Rice, D., Stilborn, H., Rowe, D., Lott, B., Branton, S., and Simmons, J. (2003).** Total and water-soluble phosphorus in broiler litter over three flocks with alum litter treatment and dietary inclusion of high available phosphorus corn and phytase supplementation. *Poult. Sci.*, 82:1544-1549.

- Nnenna, O., Emeka, N., and Okpoko, C. (2006).** Performance of broiler chicks (*Gallus domesticus*) fed maize offal-based diets supplemented with roxazyme G enzyme. *Int. J. Poult. Sci.*, 5(7): 607-610.
- NRC, National Research Council (1994).** Nutrient requirements of poultry. National Academy Press. Washington, DC., 9th Revised Edition, 19-34, USA.
- Olukosi, A., Cowienson, A., and Adeola, O. (2007).** Age-related influence of a cocktail of xylanase, amylase, and protease or phytase individually or in combination in broilers. *Poult. Sci.*, 86:77–86.
- Oluyinka, A., and Adeola, O. (2008).** Whole body accretion, growth performance and total tract nutrient retention responses of broilers to supplementation of xylanase and phytase individually or in combination in wheat-soybean meal based diets. *The Journal of Poult. Sci.*, 45: 192-198.
- Onyango, E., Bedford, M., and Adeola, O. (2005).** Efficacy of an evolved *Escherichia Coli* phytase in diets of broiler chicks. *Poult. Sci.*, 84: 248-255.
- Owens, B., Tucker, L., Collins, M., and McCracken K. (2008).** Effects of different feed additives alone or in combination on broiler performance, gut microflora and ileal histology. *Brit. Poult. Sci.*, 49 (2): 202-212.
- Parsippany, N. (2008).** Non-starch polysaccharide enzymes for poultry. Proc. 6th MIDatlantic Nutrition Conference, Univ. Maryland.

- Pettersson, D., and Åman, P. (2007).** Enzyme supplementation of a poultry diet containing rye and wheat. *Brit. J. Nutr.*, 62(1):139-149.
- Pillai, P., Connor-Dennie, T., Owens, C., and Emmert, J. (2006).** Efficacy of an *Escherichia coli* phytase in broilers fed an adequate or reduced phosphorus diets and its effect on carcass characteristics. *Poult. Sci.*, 85:1737-1745.
- Pirgozliev, V., Oduguwa, O., Acamovic, T., and Bedford, M. (2007).** Diets containing *Escherichia coli*-derived phytase on young chickens and turkeys: effects on performance, metabolizable energy, endogenous secretions, and intestinal morphology. *Poult. Sci.*, 86(4): 705-713.
- Powell, Syrena (2005).** The effect of dietary phosphorus level and phytase Supplementation on growth performance, bone breaking strength, and phosphorus excretion in broilers. M. Sc. Thesis, Graduate Faculty of the Louisiana State University and Agricultural and Mechanical College, USA.
- Qian, H., Veit, H., Komegay, E., Ravindran, V., and Denbow, D. (1996).** Effects of supplemental phytase and phosphorus on histological and other tibial bone characteristics and performances of broilers fed semi-purified diets. *Poult. Sci.*, 75:618–626.
- Qota, E., Ghamry, E., and El-Mallah, G. (2002).** Nutritive value of soaked linseed cake as affected by phytase, Biogen supplementation or formulating diets based on available amino acid on broiler performance. *Egypt. Poult. Sci.*, 22:461-475.
- Ravindran, V., Bryden, W., and Kornegay, E. (1995).** Phytates: occurrence, bioavailability and implications in poultry nutrition. *Poult. and Avian Biology Review*, 6 (2): 125-143.

- Ravindran, V., Cowieson, A., and Selle, P. (2008).** Influence of dietary electrolyte balance and microbial phytase on growth performance, nutrient utilization, and excreta quality of broiler chickens. *Poult. Sci.*, 87:677-688.
- Ravindran, V., Selle, P., and Bryden, W. (1999).** Effects of phytase supplementation, individually and in combination, with glycanase, on the nutritive value of wheat and barley. *Poult. Sci.*, 78(11): 1588-1595.
- Ravindran, V., Cabahug, S., Ravindra, G., Selle, P., and Bryden, W. (2000).** Response of broiler chickens to microbial phytase supplementation as influenced by dietary phytic acid and non-phytate phosphorous levels. II. Effects on apparent metabolisable energy, nutrient digestibility and nutrient retention. *Brit. Poult. Sci.*, 41(2):193-200.
- Ravindran, V., Morel, P., Partridge, G., Hruby, M., and Sands, J. (2006).** Influence of an *Escherichia coli*-derived phytase on nutrient utilization in broiler starters fed diets containing varying concentrations of phytic acid. *Poult. Sci.*, 85:82–89.
- Raw, S., Reddy, V., and Reddy, V. (1999).** Enhancement of phytate phosphorus availability in the diets of commercial broilers and layers. *Anim. Feed Sci. and Technol.*, 79: 211-222.
- Reitman, S., and Frankel, S. (1957).** A calorimetric methods for determination of glutamic oxaloacetic and glutamic pyruvic transaminase. *Amer. J. Clin. Pathol.*, 28:56-63

- Saleh, F., Ohtsuka, A., and Hayashi, K. (2005).** Effect of dietary enzymes on the ileal digestibility and abdominal fat content in broilers. *Anim. Sci. J.*, 76:475-478.
- Salem, F., El-Alaily, H., El-Medany, N., and Abd El-Galil, K. (2003)** Improving phosphorus utilization in broiler chick diets to minimize phosphorus pollution. *Egypt. Poult. Sci.*, 23:201-218.
- SAS. (1990).** SAS/STAT[®] User's Guide: Statistics (Release 6.04 Ed).
SAS Institute Inc. Cary, NC, USA.
- Scheideler, S., and Ferket, P. (2000).** Phytase in broiler rations: Effect on carcass yields and incidence of tibial dyschondroplasia. *J. Appl. Poult. Res.*, 9:468-475.
- Scott, T. A. (2005).** The impact of pelleting and enzyme supplementation on feed value of twenty-five Canadian wheat samples. *Proc. 17th Australian Poultry Science Symposium.*
- Sebastian, S., Touchburn, S., Chavez, E., and Lague, P. (1996).** The effects of supplemental phytase on the performance and utilization of dietary calcium, phosphorus, copper and zinc in broiler chickens fed corn-soybean diets. *Poult. Sci.*, 75: 729-736.
- Selim, Walaa (2007).** Effect of two sources of microbial phytase on productive performance meat quality and dietary protein and energy utilization by Sasso chickens. M. Sc. Thesis Faculty of Agriculture-Damanhour, Alexandria University, Egypt.
- Selle, P., and Ravindran, V. (2007).** Microbial phytase in poultry nutrition. *Anim. Feed Sci. Techno.*, 135:1–41.

- Sohail, S., and Roland, D. (1999).** Influence of supplemental phytase on performance of broilers four to six weeks of age. *Poult. Sci.*, 78(4): 550-555.
- Steel, R., and Torrie, J. (1960).** Principles and Procedures of Statistics. McGraw Hill Book Co. New York, USA.
- Steenfeldt, S., Mullertz, A., and Jensen, F. (1998).** Enzyme supplementation of wheat-based diets for broilers. 1. Effect on growth performance and intestinal viscosity *Anim. Feed Sci. and Technol.*, 75: 27-43.
- Trinder, P. (1969).** Determination of glucose in blood using oxidase with an alternative oxygen acceptor. *Ann. Clin. Biochem.*, 6:24.
- Van der Klis, J., Kwakernaak, C., and De Wit, W. (1995).** Effects of endoxylanase addition to wheat based diets on physico-chemical chyme conditions and mineral absorption in broilers. *Anim. Feed Sci. and Technol.*, 51:15-27.
- Veldman A., and Vahl, H. (1994).** Xylanase in broiler diets with differences in characteristics and content of wheat. *Brit. Poult. Sci.*, 35: 537-550.
- Vetési, M., Mézes, M., Baskay, G., and Gelencsér, E. (1998).** Effects of phytase supplementation on calcium and phosphorus output, production traits and mechanical stability of the tibia in broiler chickens. *Acta Vet. Hung.*, 46 (2): 231-242.
- Viveros, A., Brenes, A., Arija, I., and Centeno, C. (2002).** Effects of microbial phytase supplementation on mineral utilization and serum enzyme activities in broiler chicks fed different levels of phosphorus. *Poult. Sci.*, 81(8): 1172-1183

- Waldroup, P.W., Kersey, J., Saleh, E., Fritts, C., Yan, F., Stilborn, H., Crum Jr., and Raboy, V. (2000).** Nonphytate phosphorus requirement and phosphorus excretion of broiler chicks fed diets composed of normal or high available phosphorus corn with and without microbial phytase. *Poult. Sci.*, 79: 1451-1459.
- Wang, Z., Qiao, S., Lu, W., and Li, D. (2005).** Effects of enzyme supplementation on performance, nutrient digestibility, gastrointestinal morphology, and volatile fatty acid profiles in the hindgut of broilers fed wheat-based diets. *Poult. Sci.*, 84(6): 875.
- Watson, D. (1960).** Determination of cholesterol in blood. *Clinical Chemistry Acta*, 5: 637.
- Yan, F., Kersey, J., Fritts, C., Waldroup, P., Stilborn, H., Crumm, R., Rice, D., and Raboy, V. (2000).** Evaluation of normal yellow dent corn and high available phosphorus corn in combination with reduced dietary phosphorus and phytase supplementation for broilers grown to market weight in litter pens. *Poult. Sci.*, 79:1282-1289.
- Zanella I., Sakomura, N. K., Silversides, F. G., Figueirido, A., and Pack, M. (1999).** Effect of enzyme supplementation of broiler diets based on corn and soybeans. *Poult. Sci.*, 78(4): 561.
- Zanini, S., and Sazzad, M. (1999).** Effects of microbial phytase on growth and mineral utilization in broilers fed on maize soybean-based diets. *Brit. Poult. Sci.*, 40:348–352.

Zhou, Y., Jiang, Z., Lv, D., and Wang, T. (2009). Improved energy-utilizing efficiency by enzyme preparation supplement in broiler diets with different metabolizable energy levels. *Poult. Sci.*, 88: 316–322.

Zollner, N., and Kirsch, K. (1962). Test combination. Total lipids. Sulfo-phospho-vanillin reaction. *Furdie Gesampte Exp. Med.*, 135: 545.

Zyła, K., Wikiera, A., Koreleski, J., Swiatkiewicz, S., Piironen, J., and Ledoux, D. (2000). Comparison of the efficacies of a novel *Aspergillus niger* mycelium with separate and combined effectiveness of phytase, acid phosphatase, and pectinase in dephosphorylation of wheat-based feeds fed to growing broilers. *Poult. Sci.*, 79(10):1434-1443.

7. APPENDIX

Live weight gain

General Linear Models Procedure

Dependent Variable: W0

Source	DF	Sum of Squares	Mean Square	F Value	Pr > F
Model	7	0.36468750	0.05209821	0.46	0.8528
Error	24	2.71250000	0.11302083		
Corrected Total	31	3.07718750			
	R-Square	C.V.	Root MSE		W0 Mean
	0.118513	0.835050	0.33618571		40.25937500
Source	DF	Type I SS	Mean Square	F Value	Pr > F
ME	1	0.00781250	0.00781250	0.07	0.7949
ENZ	3	0.23593750	0.07864583	0.70	0.5637
INT	3	0.12093750	0.04031250	0.36	0.7847

Dependent Variable: Weight gain (0-3 wks)

Source	DF	Sum of Squares	Mean Square	F Value	Pr > F
Model	7	32115.56468750	4587.93781250	15.93	0.0001
Error	24	6911.11250001	287.96302083		
Corrected Total	31	39026.67718751			
	R-Square	C.V.	Root MSE		WGS Mean
	0.822913	2.104194	16.96947320		806.45937500
Source	DF	Type I SS	Mean Square	F Value	Pr > F
ME	1	23355.00781250	23355.00781250	81.10	0.0001
ENZ	3	4045.78593750	1348.59531250	4.68	0.0103
INT	3	4714.77093750	1571.59031250	5.46	0.0053

Dependent Variable: Weight gain (4-6 wks)

Source	DF	Sum of Squares	Mean Square	F Value	Pr > F
Model	7	413244.46875000	59034.92410714	44.23	0.0001
Error	24	32030.75000000	1334.61458333		
Corrected Total	31	445275.21875000			
	R-Square	C.V.	Root MSE		WGF Mean
	0.928065	2.507961	36.53237719		1456.65625000
Source	DF	Type I SS	Mean Square	F Value	Pr > F
ME	1	182257.03125000	182257.03125000	136.56	0.0001
ENZ	3	80600.84375000	26866.94791667	20.13	0.0001
INT	3	150386.59375000	50128.86458333	37.56	0.0001

Dependent Variable: WG (0-6 wks)

Source	DF	Sum of Squares	Mean Square	F Value	Pr > F
Model	7	536111.83968750	76587.40566964	30.83	0.0001
Error	24	59627.96250005	2484.49843750		
Corrected Total	31	595739.80218755			
	R-Square	C.V.	Root MSE		WGT Mean
	0.899909	2.202483	49.84474333		2263.11562500
Source	DF	Type I SS	Mean Square	F Value	Pr > F
ME	1	336097.50781250	336097.50781250	135.28	0.0001
ENZ	3	58488.06093750	19496.02031250	7.85	0.0008
INT	3	141526.27093750	47175.42364583	18.99	0.0001

Feed conversion ratio

General Linear Models Procedure

Dependent Variable: FC (0-3 wks)

Source	DF	Sum of Squares	Mean Square	F Value	Pr > F
Model	7	0.11152099	0.01593157	5.06	0.0012
Error	24	0.07555567	0.00314815		
Corrected Total	31	0.18707665			
	R-Square	C.V.	Root MSE		FCS Mean
	0.596125	3.935533	0.05610840		1.42568750
Source	DF	Type I SS	Mean Square	F Value	Pr > F
ME	1	0.08618168	0.08618168	27.38	0.0001
ENZ	3	0.01947915	0.00649305	2.06	0.1319
INT	3	0.00586015	0.00195338	0.62	0.6086

Dependent Variable: FC (4-6 wks)

Source	DF	Sum of Squares	Mean Square	F Value	Pr > F
Model	7	1.22455511	0.17493644	58.89	0.0001
Error	24	0.07129950	0.00297081		
Corrected Total	31	1.29585461			
	R-Square	C.V.	Root MSE		FCF Mean
	0.944979	2.540293	0.05450516		2.14562500
Source	DF	Type I SS	Mean Square	F Value	Pr > F
ME	1	0.50635339	0.50635339	170.44	0.0001
ENZ	3	0.31306419	0.10435473	35.13	0.0001
INT	3	0.40513753	0.13504584	45.46	0.0001

Dependent Variable: FC (0-6 wks)

Source	DF	Sum of Squares	Mean Square	F Value	Pr > F
Model	7	0.43466520	0.06209503	28.86	0.0001
Error	24	0.05163601	0.00215150		
Corrected Total	31	0.48630122			
	R-Square	C.V.	Root MSE	FCT Mean	
	0.893819	2.597604	0.04638427	1.78565625	
Source	DF	Type I SS	Mean Square	F Value	Pr > F
ME	1	0.25258278	0.25258278	117.40	0.0001
ENZ	3	0.08421927	0.02807309	13.05	0.0001
INT	3	0.09786316	0.03262105	15.16	0.0001

Carcass characteristics

General Linear Models Procedure

Dependent Variable: CARCASS

Source	DF	sum of Squares	Mean Square	F Value	Pr > F
Model	7	22.14286667	3.16326667	1.90	0.0953
Error	40	66.63793333	1.66594833		
Corrected Total	47	88.78080000			
	R-Square	Coeff Var	Root MSE	CARCASS Mean	
	0.249411	1.768468	1.290716	72.98500	
Source	DF	Type I SS	Mean Square	F Value	Pr > F
ME	1	5.24040833	5.24040833	3.15	0.0837
ENZ	3	13.79295000	4.59765000	2.76	0.0546
ME*ENZ	3	3.10950833	1.03650278	0.62	0.6048

Dependent Variable: LIVER

Source	DF	Sum of Squares	Mean Square	F Value	Pr > F
Model	7	0.95923333	0.13703333	4.29	0.0013
Error	40	1.27826667	0.03195667		
Corrected Total	47	2.23750000			
	R-Square	Coeff Var	Root MSE	LIVER Mean	
	0.428708	9.226543	0.178764	1.937500	
Source	DF	Type I SS	Mean Square	F Value	Pr > F
ME	1	0.08333333	0.08333333	2.61	0.1142
ENZ	3	0.78151667	0.26050556	8.15	0.0002
ME*ENZ	3	0.09438333	0.03146111	0.98	0.4098

General Linear Models Procedure

Dependent Variable: HEART

Source	DF	Sum of Squares	Mean Square	F Value	Pr > F
Model	7	0.07193125	0.01027589	0.97	0.4635
Error	40	0.42211667	0.01055292		
Corrected Total	47	0.49404792			

R-Square	Coeff Var	Root MSE	HEART Mean
0.145596	18.98696	0.102727	0.541042

Source	DF	Type I SS	Mean Square	F Value	Pr > F
ME	1	0.00025208	0.00025208	0.02	0.8779
ENZ	3	0.05010625	0.01670208	1.58	0.2086
ME*ENZ	3	0.02157292	0.00719097	0.68	0.5686

Dependent Variable: GIZZARD

Source	DF	Sum of Squares	Mean Square	F Value	Pr > F
Model	7	0.60404792	0.08629256	1.98	0.0819
Error	40	1.74215000	0.04355375		
Corrected Total	47	2.34619792			

R-Square	Coeff Var	Root MSE	GIZZARD Mean
0.257458	14.44050	0.208695	1.445208

Source	DF	Type I SS	Mean Square	F Value	Pr > F
ME	1	0.05535208	0.05535208	1.27	0.2663
ENZ	3	0.35787292	0.11929097	2.74	0.0559
ME*ENZ	3	0.19082292	0.06360764	1.46	0.2398

Dependent Variable: FAT

Source	DF	Sum of Squares	Mean Square	F Value	Pr > F
Model	7	0.82786458	0.11826637	2.11	0.0650
Error	40	2.24541667	0.05613542		
Corrected Total	47	3.07328125			

R-Square	Coeff Var	Root MSE	FAT Mean
0.269375	10.97212	0.236929	2.159375

Source	DF	Type I SS	Mean Square	F Value	Pr > F
ME	1	0.10546875	0.10546875	1.88	0.1781
ENZ	3	0.58432292	0.19477431	3.47	0.0248
ME*ENZ	3	0.13807292	0.04602431	0.82	0.4906

Blood constituents

General Linear Models Procedure

Dependent Variable: Total Protein

Source	DF	Sum of Squares	Mean Square	F Value	Pr > F
Model	7	5.5100000	0.78714286	2.84	0.0263
Error	24	6.6500000	0.27708333		
Corrected Total	31	12.1600000			
	R-Square	C.V.	Root MSE		TP Mean
	0.453125	10.12283	0.52638706		5.2000000
Source	DF	Type I SS	Mean Square	F Value	Pr > F
ME	1	2.10125000	2.10125000	7.58	0.0111
ENZ	3	0.64250000	0.21416667	0.77	0.5205
INT	3	2.76625000	0.92208333	3.33	0.0365

General Linear Models Procedure

Dependent Variable: Albumin

Source	DF	Sum of Squares	Mean Square	F Value	Pr > F
Model	7	0.76968750	0.10995536	1.26	0.3090
Error	24	2.08750000	0.08697917		
Corrected Total	31	2.85718750			
	R-Square	C.V.	Root MSE		ALB Mean
	0.269386	9.699398	0.29492231		3.04062500
Source	DF	Type I SS	Mean Square	F Value	Pr > F
ME	1	0.05281250	0.05281250	0.61	0.4435
ENZ	3	0.09593750	0.03197917	0.37	0.7770
INT	3	0.62093750	0.20697917	2.38	0.0947

General Linear Models Procedure

Dependent Variable: Globulin

Source	DF	Sum of Squares	Mean Square	F Value	Pr > F
Model	7	2.47500000	0.35357143	4.24	0.0036
Error	24	2.00000000	0.08333333		
Corrected Total	31	4.47500000			
	R-Square	C.V.	Root MSE		GLOB Mean
	0.553073	13.19658	0.28867513		2.18750000
Source	DF	Type I SS	Mean Square	F Value	Pr > F
ME	1	1.20125000	1.20125000	14.42	0.0009
ENZ	3	0.45250000	0.15083333	1.81	0.1723
INT	3	0.82125000	0.27375000	3.29	0.0381

General Linear Models Procedure

Dependent Variable: Albumin/Globulin ratio

Source	DF	Sum of Squares	Mean Square	F Value	Pr > F
Model	7	0.46820000	0.06688571	3.37	0.0119
Error	24	0.47595000	0.01983125		
Corrected Total	31	0.94415000			
	R-Square	C.V.	Root MSE		AGR Mean
	0.495896	10.10393	0.14082347		1.39375000
Source	DF	Type I SS	Mean Square	F Value	Pr > F
ME	1	0.27380000	0.27380000	13.81	0.0011
ENZ	3	0.14122500	0.04707500	2.37	0.0953
INT	3	0.05317500	0.01772500	0.89	0.4586

General Linear Models Procedure

Dependent Variable: Glucose

Source	DF	Sum of Squares	Mean Square	F Value	Pr > F
Model	7	10108.37500000	1444.05357143	4.12	0.0042
Error	24	8411.50000000	350.47916667		
Corrected Total	31	18519.87500000			
	R-Square	C.V.	Root MSE		GLU Mean
	0.545812	16.14757	18.72108882		115.93750000
Source	DF	Type I SS	Mean Square	F Value	Pr > F
ME	1	3960.50000000	3960.50000000	11.30	0.0026
ENZ	3	855.62500000	285.20833333	0.81	0.4988
INT	3	5292.25000000	1764.08333333	5.03	0.0076

General Linear Models Procedure

Dependent Variable: Triglycerides

Source	DF	Sum of Squares	Mean Square	F Value	Pr > F
Model	7	223.71875000	31.95982143	1.97	0.1023
Error	24	389.75000000	16.23958333		
Corrected Total	31	613.46875000			
	R-Square	C.V.	Root MSE		TRI Mean
	0.364678	4.620379	4.02983664		87.21875000
Source	DF	Type I SS	Mean Square	F Value	Pr > F
ME	1	81.28125000	81.28125000	5.01	0.0348
ENZ	3	64.34375000	21.44791667	1.32	0.2908
INT	3	78.09375000	26.03125000	1.60	0.2148

General Linear Models Procedure

Dependent Variable: Cholesterol

Source	DF	Sum of Squares	Mean Square	F Value	Pr > F
Model	7	796.71875000	113.81696429	1.17	0.3583
Error	24	2343.75000000	97.65625000		
Corrected Total	31	3140.46875000			
	R-Square	C.V.	Root MSE		CHOL Mean
	0.253694	12.78721	9.88211769		77.28125000

Source	DF	Type I SS	Mean Square	F Value	Pr > F
ME	1	215.28125000	215.28125000	2.20	0.1506
ENZ	3	465.09375000	155.03125000	1.59	0.2184
INT	3	116.34375000	38.78125000	0.40	0.7563

General Linear Models Procedure

Dependent Variable: Creatinine

Source	DF	Sum of Squares	Mean Square	F Value	Pr > F
Model	7	0.07538750	0.01076964	3.39	0.0116
Error	24	0.07620000	0.00317500		
Corrected Total	31	0.15158750			
	R-Square	C.V.	Root MSE		CRE Mean
	0.497320	27.57047	0.05634714		0.20437500

Source	DF	Type I SS	Mean Square	F Value	Pr > F
ME	1	0.01201250	0.01201250	3.78	0.0636
ENZ	3	0.03206250	0.01068750	3.37	0.0352
INT	3	0.03131250	0.01043750	3.29	0.0380

General Linear Models Procedure

Dependent Variable: ALT

Source	DF	Sum of Squares	Mean Square	F Value	Pr > F
Model	7	8.16375000	1.16625000	2.36	0.0556
Error	24	11.88500000	0.49520833		
Corrected Total	31	20.04875000			
	R-Square	C.V.	Root MSE		ALT Mean
	0.407195	30.34870	0.70371040		2.31875000

Source	DF	Type I SS	Mean Square	F Value	Pr > F
ME	1	4.96125000	4.96125000	10.02	0.0042
ENZ	3	1.72625000	0.57541667	1.16	0.3448
INT	3	1.47625000	0.49208333	0.99	0.4126

General Linear Models Procedure

Dependent Variable: AST

Source	DF	Sum of Squares	Mean Square	F Value	Pr > F
Model	7	80916.71875000	11559.53125000	2.87	0.0250
Error	24	96556.25000000	4023.17708333		
Corrected Total	31	177472.96875000			

R-Square	C.V.	Root MSE	AST Mean
0.455938	30.94546	63.42851948	204.96875000

Source	DF	Type I SS	Mean Square	F Value	Pr > F
ME	1	29951.28125000	29951.28125000	7.44	0.0117
ENZ	3	16878.59375000	5626.19791667	1.40	0.2675
INT	3	34086.84375000	11362.28125000	2.82	0.0602

ملخص الدراسة

قال ﷺ: ﴿وَقُلِ اعْمَلُوا فَسَيَرَى اللَّهُ عَمَلَكُمْ وَرَسُولُهُ وَالْمُؤْمِنُونَ وَسَتُرَدُّونَ إِلَىٰ عَالَمِ الْغَيْبِ وَالشَّهَادَةِ فَيُنَبِّئُكُمْ بِمَا كُنْتُمْ تَعْمَلُونَ﴾ (1).

الحمد لله رب العالمين، الأول بلا بداية، والآخر بلا نهاية، والصلاة على من لا نبي بعده،
وبعد،

فقد هدفت الدراسة إلى مقارنة تأثير مصادر مختلفة من الأنزيمات على استخدام الطاقة الممتلة في علائق الدجاج اللاحم التي تحتوى على القمح.

ولتحقيق هدف الدراسة:

تم توزيع ثلاثمائة وعشرين كتكوتاً عمر يوم واحد، غير مجنسة من سلالة كوب عشوائياً على (8) ثماني مجموعات تجريبية بحيث كان كل (40) أربعون كتكوتاً لكل معاملة وتم إضافة (4) أربعة مصادر للأنزيم مع مستويين للطاقة الممتلة على أداء النمو، وصفات الذبيحة، وبعض مكونات مصل الدم.

وكانت العلائق المستخدمة على النحو التالي:

- *** **عليقة 1:** مستوى أمثل للطاقة الممتلة دون إضافة للأنزيم.
- *** **عليقة 2:** مستوى أمثل للطاقة الممتلة مع إضافة للأنزيم Avizyme 1500 (100 جم/100 كجم).
- *** **عليقة 3:** مستوى أمثل للطاقة الممتلة مع إضافة للأنزيم Phyzyme XP TPT 10000 (50 جم/100 كجم).
- *** **عليقة 4:** مستوى أمثل للطاقة الممتلة مع إضافة للأنزيم Avizyme 1500 (50 جم/100 كجم) + أنزيم Phyzyme XP TPT 10000 (25 جم/100 كجم).
- *** **عليقة 5:** مستوى طاقة ممتلة يقل (100) مائة كالوري/ كجم عليقة عن المستوى الامثل للطاقة الممتلة دون إضافة للأنزيم.

(1)سورة التوبة: من الآية {105}.

- *** **عليقة6:** مستوى طاقة ممثلة يقل (100)مائة كالوري/ كجم عليقة عن المستوى الأمثل للطاقة الممثلة مع إضافة لأنزيم Avizyme 1500 (100 جم/كجم 100).
 *** **عليقة7:** مستوى طاقة ممثلة يقل (100) مائة كالوري/ كجم عليقة عن المستوى الأمثل للطاقة الممثلة مع إضافة لأنزيم Phyzyme XP TPT 10000 (50 جم/كجم 100 كجم).
 *** **عليقة8:** مستوى طاقة ممثلة يقل (100)مائة كالوري/ كجم عليقة عن المستوى الأمثل للطاقة الممثلة مع إضافة لأنزيم Avizyme 1500 (50 جم/كجم 100 كجم) + أنزيم PhyzymeXP TPT 10000 (25 جم/كجم 100).

يمكن تلخيص النتائج على النحو التالي:

- مستوى الطاقة الممثلة الأمثل أفضل معنوياً من المستوى الأقل في معدل النمو والكفاءة التحويلية للغذاء.
- إضافة الأنزيمات أعطى تأثيراً إيجابياً لحدّ ما على أداء النمو والخليط بين مصدري الأنزيم أعطى أفضل أداء.
- مستوى طاقة الغذاء لم يكن له تأثير معنوي على أي من صفات الذبيحة.
- إضافة أنزيم Phyzyme XP TPT 10000 أعطى معنوياً أعلى نسبة تصافٍ للذبيحة بين مصادر الأنزيمات المستخدمة، بينما عليقة المقارنة كانت الأعلى في وزني الكبد والقانصة، والأقل في وزن دهن البطن مقارنة بإضافة الأنزيمات للعلائق.
- مستوى طاقة الغذاء الأمثل أعطى معنوياً زيادة في محتوى مصل الدم من البروتين الكلى، الجلوبيولين، الجلوكوز، الجلوسريدات الكلية، ووظائف الكبد، بينما أدى إلى انخفاض نسبة الألبومين/ الجلوبيولين مقارنةً بالمستوى الأقل للطاقة الممثلة.
- إضافة الأنزيمات لم يكن له تأثير معنوي على قياسات مصل الدم.
- مستوى طاقة الغذاء الأمثل كان أعلى في مقياس الأداء مقارنةً بالمستوى الأقل.
- إضافة أنزيم Phyzyme XP TPT 10000 كان الأفضل بالنسبة لمقياس الأداء.

التوصيات :-

من نتائج الدراسة نخلص إلى أنه يمكن إضافة مخاليط الإنزيمات إلى علائق الدواجن منخفضة المحتوى من الطاقة الممتلئة لتحسين الأداء الإنتاجي، وصفات الذبيحة للدجاج اللاحم. ونوصي بتجارب أخرى لدراسة تأثير تلك الإضافات الإنزيمية على بعض المخلفات الزراعية المتوفرة في الأراضي الفلسطينية، مثل، جفت الزيتون، مسحوق نوى البلح، لتحسين الاستفادة منها، وإدخالها في تكوين علائق اقتصادية لفراريج اللحم.



جامعة القاهرة ج.م.ع

البرنامج المشترك



جامعة الأزهر - غزة

لجنة الإشراف

اسم الطالب: *وسام صادق عبدالله أبو سليمان*

عنوان الرسالة: *دراسة مقارنة لتأثير مصادر مختلفة من الإنزيمات على الاستفادة من طاقة العليقة في دجاج التسمين*

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جامعة القاهرة ج.م.ع

البرنامج المشترك



جامعة الأزهر - غزة

دراسة مقارنة

لتأثير مصادر مختلفة من الإنزيمات على الاستفادة من طاقة العليقة في دجاج التسمين

رسالة مقدمة من /

وسام صادق عبدالله أبو سليمان

بكالوريوس العلوم الزراعية (قسم الإنتاج الحيواني والدواجن)

كلية الزراعة - جامعة الأزهر - غزة - فلسطين 2007م

كجزء من متطلبات الحصول على درجة الماجستير في العلوم الزراعية
(إنتاج حيواني ودواجن)

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